

School of Artificial Intelligence Applied to Microbiomes

01-10-2025 – 03-10-2025
AgroParisTech



université
PARIS-SACLAY

GRADUATE SCHOOL
Biosphera



ÉCOLE DOCTORALE
Sciences du végétal:
du gène à l'écosystème
(SEVE)

METAPROGRAMME
HOLOFLUX

MicroEngine project



Programme EXPLOR'AE



Special thanks to

Daniel Garza for organizing the school, as well as
Aristeidis Litos and all the speakers

Frédérique Delville from Biosphera

THEMATIC PROGRAM

MICROORGANISMS: FROM GENES TO ECO-SYSTEMS

Miguel Iniesto (IDEEV, ESE, miguel.iniesto@universite-paris-saclay.fr)

Ariane Bize (INRAE, PROSE, ariane.bize@inrae.fr)

aims to reinforce the network of Paris-Saclay's students and scientists working in microbiology, at a broad sense, on Biosphera topics

meeting once a year

financial support of collective events related to the thematic program → don't hesitate to contact us!

Journée Scientifique Biosphera

Jeudi 15 janvier 2026 de 9h à 19h – Campus Agro Paris-Saclay

Elle aura pour thématique **Biodiversité & évolution à l'épreuve du changement climatique** »

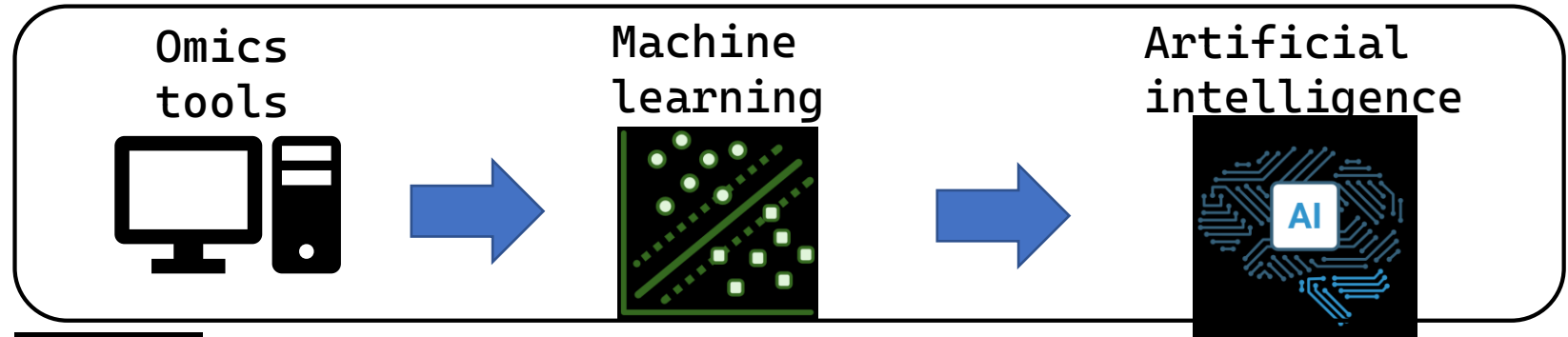
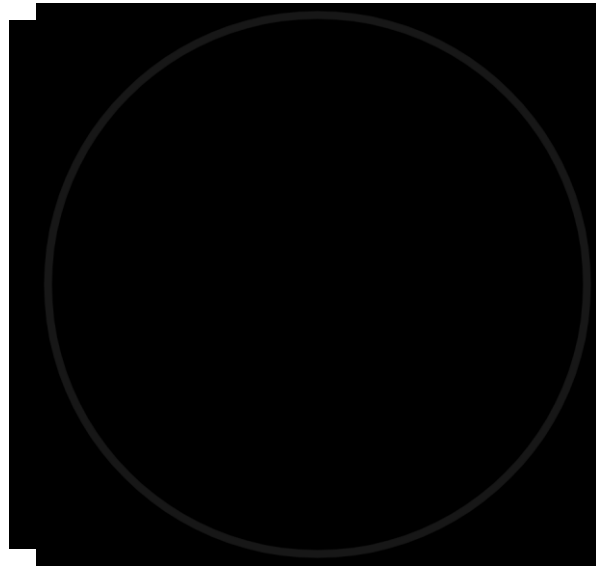
Le programme est en cours de construction. Il vous sera transmis sous peu avec le lien d'inscription.

School overview & practical information

Daniel Garza



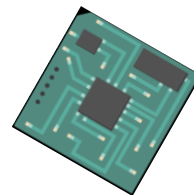
Goal of the School: provide a broad overview of how advanced computational methods, including artificial intelligence, are used in microbiome science



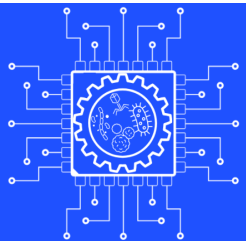
New ways to make sense of large microbiome datasets



New models to predict and explain microbiome dynamics



New approaches to engineering microbial systems for targeted functions or desired community states



**SCHOOL OF ARTIFICIAL
INTELLIGENCE APPLIED TO
MICROBIOMES**

Program: research talks & tutorials



New ways to make sense of large microbiome datasets



Dr. Julian Tap

INRAE

France

Lecture: *Integrating Human Gut Microbiome Data at Scale: From Individuals to Multi-Layered Features in Clinical and Nutritional Studies*

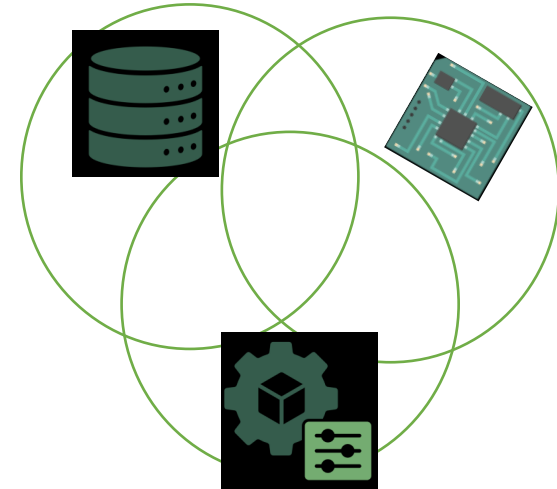


Prof. Bas Dutilh

Friedrich Schiller University Jena

Germany

Lecture: *Mapping the Microverse and Modelling its Drivers*



Program: research talks & tutorials



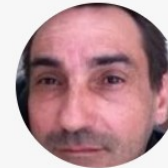
New computational models to predict and explain microbiome dynamics



Dr. Clémence Frioux

Inria, University of Bordeaux
France

***Lecture:** to be defined*



Prof. Jean-Loup Faulon

University of Paris Saclay, INRAE
France

***Lecture:** Bacterial Reservoir Computing*

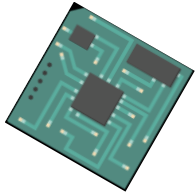


Dr. Haris Zafeiropoulos

KU Leuven
Belgium

***Lecture:** to be defined*

Program: research talks & tutorials



New approaches to engineering microbial systems for targeted functions or desired community states



Dr. Djordje Bajić

TU Delft
The Netherlands

Lecture: *Statistical mapping of Structure-Function Landscapes in Microbiomes*



Dr. Lucas Böttcher

Frankfurt School of Finance & Management
Germany

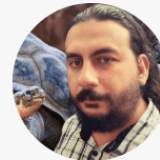
Lecture: *Simulation and Control of High-Dimensional Dynamical Systems Using Artificial Neural Networks*



Dr. Alex Fedorec

University College London
United Kingdom

Lecture: *Computational Design of Synthetic Microbial Communities*



Dr. Daniel Garza

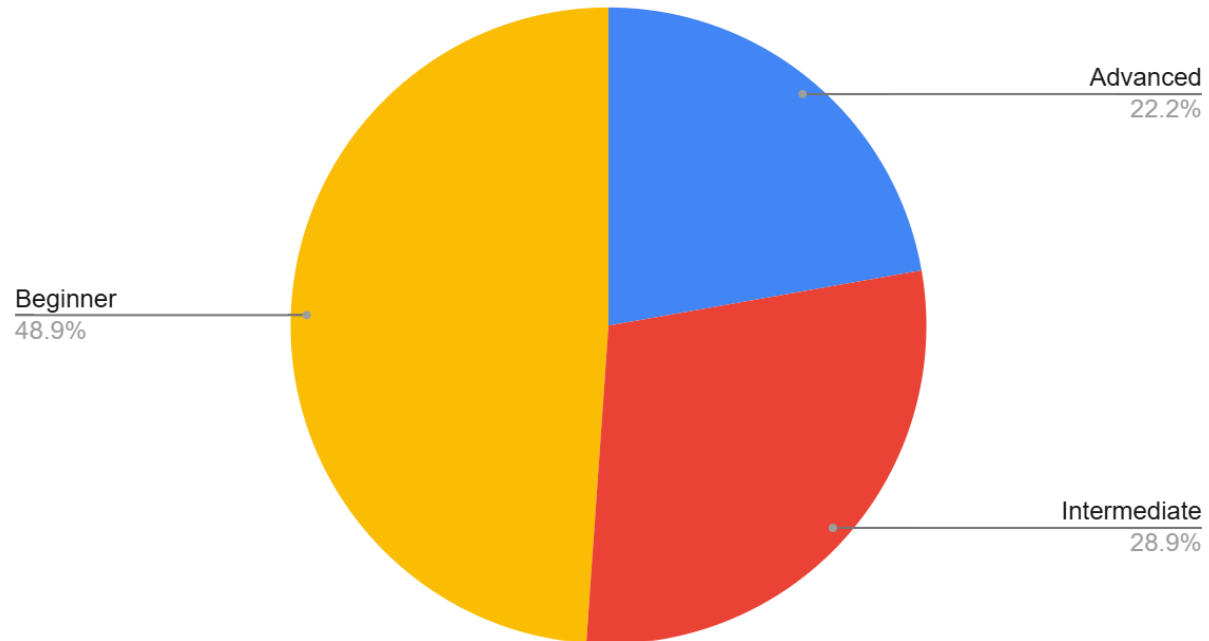
University of Paris Saclay, INRAE
France

Lecture: *Controlling complex microbiomes with computational intelligence: physical and virtual prototypes*

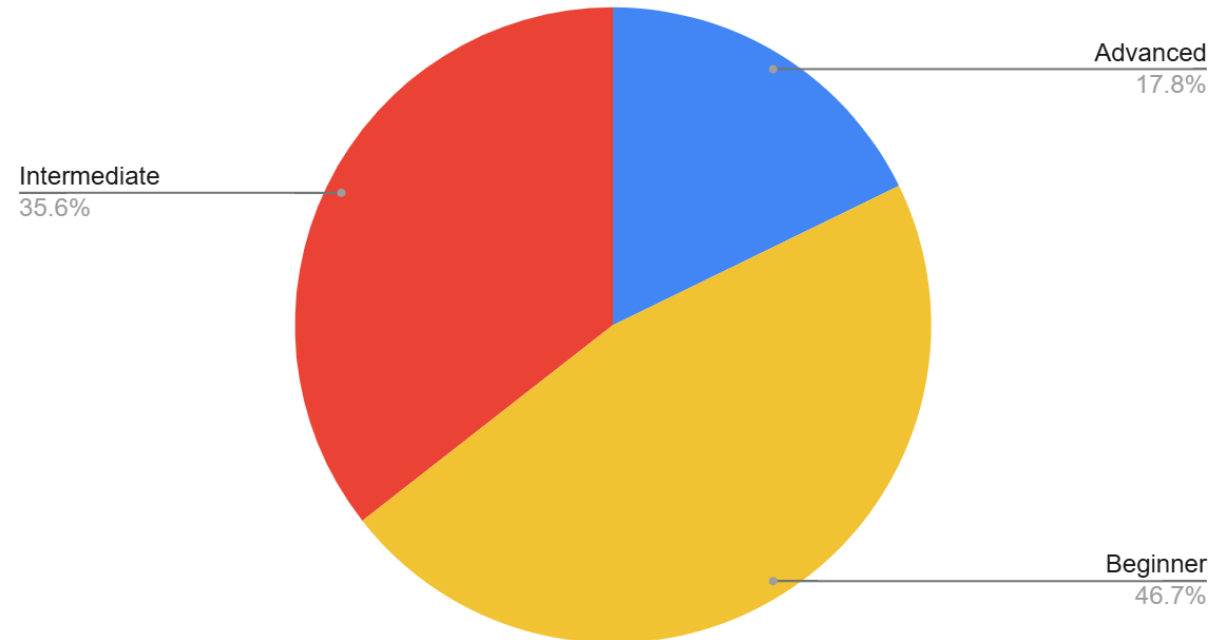
Participants

48 registrations

Python Level



R level



General Information

Program/Speakers/Participants:

<https://ai-microbiome-school.onrender.com/>

Slides and materials will be later organized in the “Tutorial & Slides” section of the website.

Most of the code will be available in the GitHub repository:

<https://github.com/danielriosgarza/AiSchool.git>

IFB Cluster



INSTITUT FRANÇAIS DE BIOINFORMATIQUE

<https://ondemand.cluster.france-bioinformatique.fr>



INSTITUT FRANÇAIS DE BIOINFORMATIQUE

Sign in to access your account

Sign in

Lost your password? [RESET](#)

Grant Access

Open OnDemand would like to:

- View basic profile information
 - View your email address
 - View your groups
-

Grant Access

Cancel

Clusters ▼

Interactive Apps ▼



>_ Core cluster Shell Access

Warning: Permanently added 'core-login1.cluster.france-bioinformatique.fr,192.168.16.205' (ECDSA) to the list of known hosts.
tp182674@core-login1.cluster.france-bioinformatique.fr's password:

Welcome to the

IFB Core Cluster

■ Documentation: <https://doc.cluster.france-bioinformatique.fr>
⚙ Cluster Support: <https://support.cluster.france-bioinformatique.fr>
🔧 Community Forum: <https://community.france-bioinformatique.fr>

⚠ ATTENTION MAINTENANCE 6-10 Octobre 2025 🚧
Arrêt du Cluster le Lundi 6 octobre à 14h
Reprise du service le Vendredi 10 Octobre à 18h

Last login: Sat Sep 27 14:39:52 2025 from 192.168.16.235

HOME [#-----] 7 / 100 GB

Update: 2025-09-29 17:00 - Your current default account is tp_2534_ai_microbiomes_181502 - More info: statusBars --help
tp182674@clust-slurm-client:~\$


```
tp182674@clust-slurm-client:~$ git clone https://github.com/danielriosgarza/AiSchool.git
```

git clone https://github.com/danielriosgarza/AiSchool.git

```
tp182674@clust-slurm-client:~$ git clone https://github.com/danielriosgarza/AiSchool.git
Cloning into 'AiSchool'...
remote: Enumerating objects: 506, done.
remote: Counting objects: 100% (98/98), done.
remote: Compressing objects: 100% (69/69), done.
remote: Total 506 (delta 61), reused 57 (delta 27), pack-reused 408 (from 1)
Receiving objects: 100% (506/506), 54.14 MiB | 39.26 MiB/s, done.
```

OPEN

OnDemand

OnDemand provides an integrated, single access point for all of your HPC resources.

Pinned Apps A featured subset of [all available apps](#)



JupyterLab: Core
System Installed App



RStudio Server: Core
System Installed App



RTrainer: Core
System Installed App



Desktop: Core
System Installed App



Reservation

2534_AI_microbiomes



Account

tp_2534_ai_microbiomes_181502



Partition

fast



Number of CPUs

3

A least one CPU is required for the JupyterLab webserver

Amount of memory

10G

Unit can be specified using the suffix M, G or T (default is megabytes).

GPUs

No GPU



Number of GPUs

0

Number of hours

3

Launch

JupyterLab: Core (61720581)

1 node

3 cores

Running

Host: cpu-node-102

 Delete

Created at: 2025-09-29 17:26:36 CEST

Time Remaining: 2 hours and 59 minutes

Session ID: e18d72b7-da0c-427f-8b03-d66f8d38156b

Reservation: 2534_AI_microbiomes

Account: tp_2534_ai_microbiomes_181502

Partition: fast

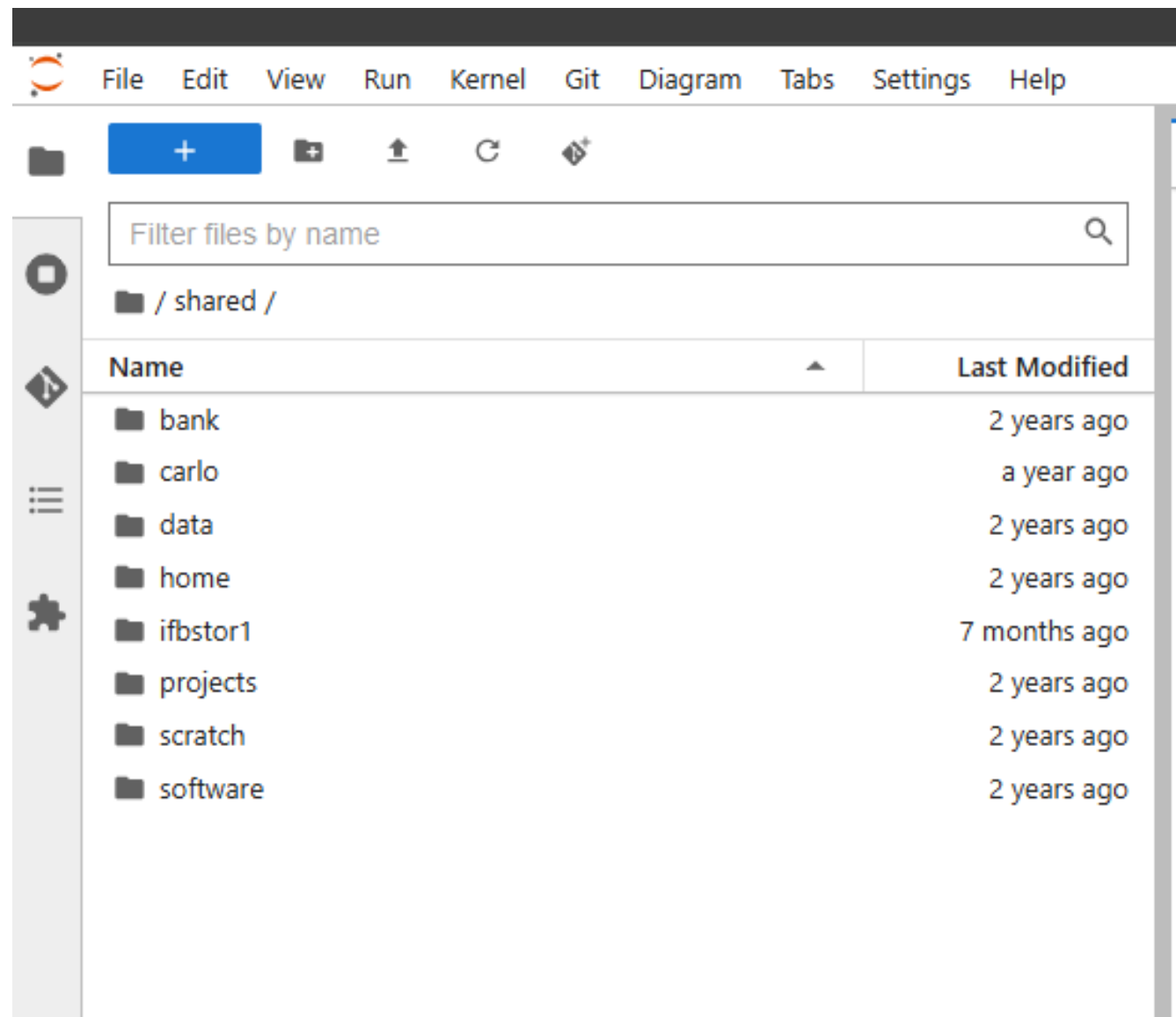
Number of CPUs: 3

Amount of memory: 10G

GPUs:

Number of GPUs: 0

  Connect to Jupyter



Go to: `shared/home/<tp user name>/`
`AiSchool/content/DanielGarza/notebooks/pip_install.ipynb`

Select Kernel

Select kernel for: "pip_install.ipynb"

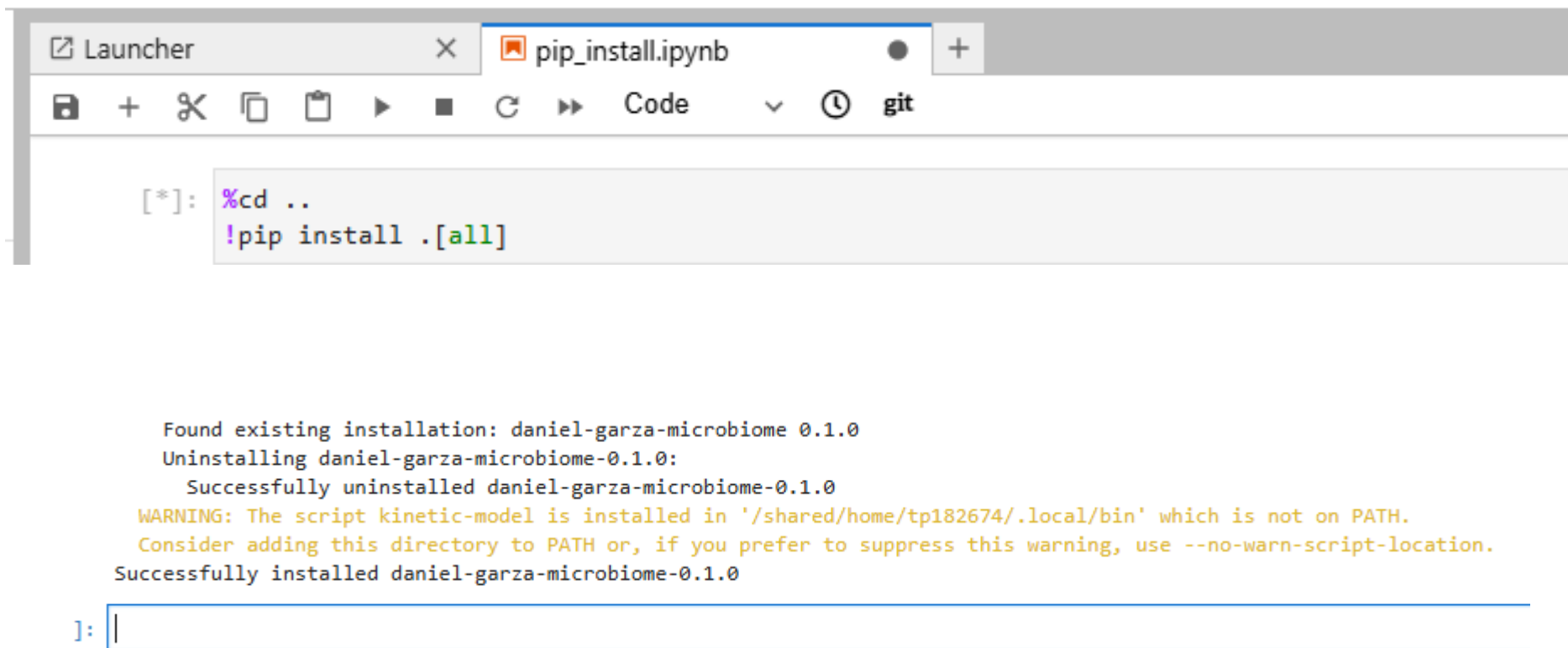
Python 3.12



No Kernel

Select

Run “play button” or “shift + enter”



The screenshot shows a Jupyter Notebook window with a tab titled 'pip_install.ipynb'. The interface includes a top bar with a 'Launcher' button, a close button, and a '+' button. Below the top bar is a toolbar with icons for saving, adding, deleting, copying, pasting, running (play button), and other actions. The main area contains a code cell with the following text:

```
[*]: %cd ..  
      !pip install .[all]
```

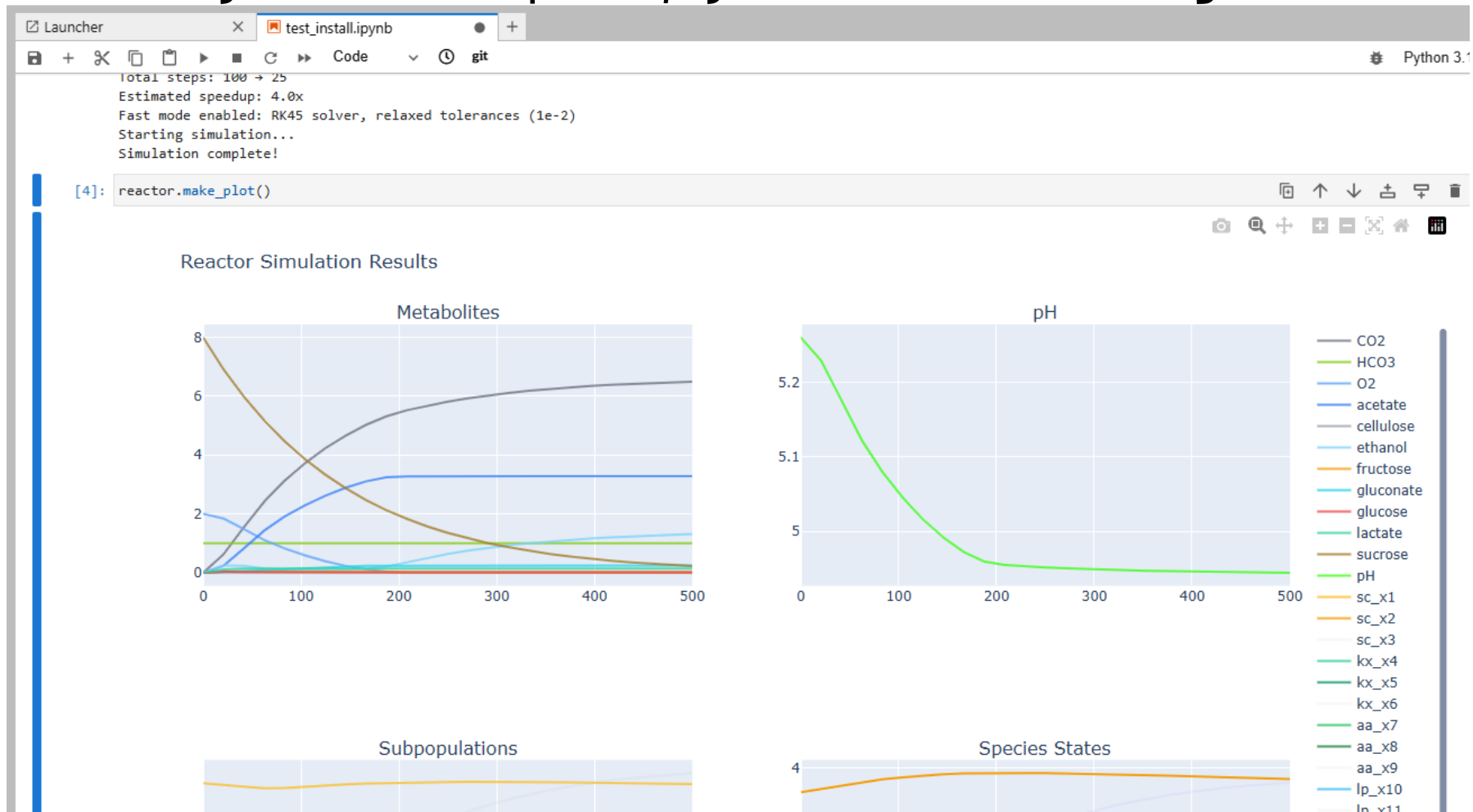
The output of the code cell is displayed below the input area:

```
Found existing installation: daniel-garza-microbiome 0.1.0  
Uninstalling daniel-garza-microbiome-0.1.0:  
  Successfully uninstalled daniel-garza-microbiome-0.1.0  
WARNING: The script kinetic-model is installed in '/shared/home/tp182674/.local/bin' which is not on PATH.  
Consider adding this directory to PATH or, if you prefer to suppress this warning, use --no-warn-script-location.  
Successfully installed daniel-garza-microbiome-0.1.0
```

At the bottom of the code cell, there is a prompt ']:' followed by a blue-bordered input box containing a vertical cursor.

Open and run the notebook “test_install.ipynb

Wait a min... if you see the plots, you are all set to go.

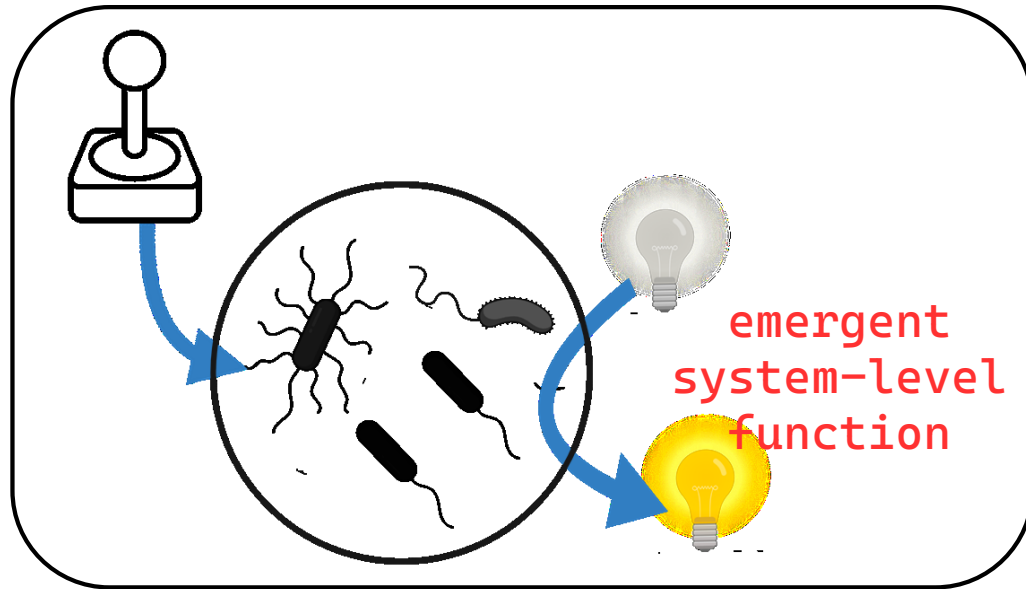


Controlling complex microbiomes with computational intelligence: physical and virtual prototypes

Daniel Garza



Microbiome engineering is the rational control of microbial ecosystems to make a desired collective function emerge and persist.

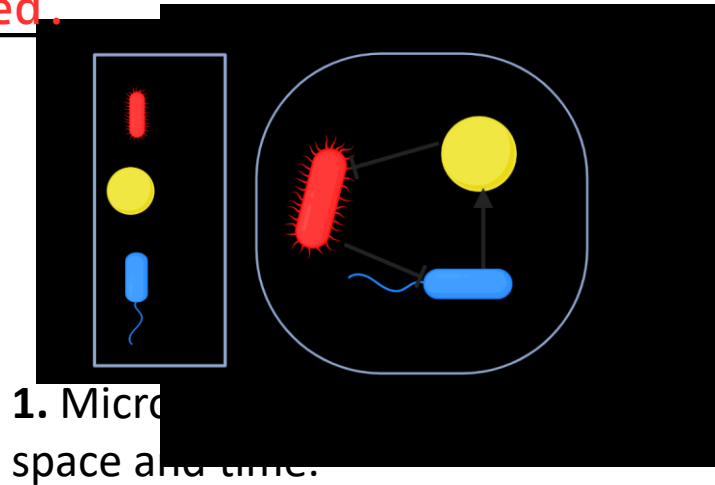


However, there are some important problems...

Problem #1: microbiomes are difficult to predict

One way uncover a system's dynamics is to model it. But classical modeling assumes that parameters are fixed and identifiable.

For example, the generalized Lotka-Volterra model, widely used in ecology, is built on the premise that growth rates and interaction strengths are fixed and can be uniquely identified.

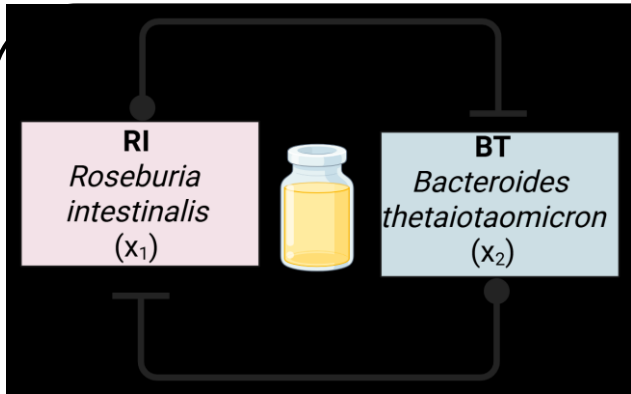


	A	B	C
A	-1.00	0.00	-0.05
B	-0.10	-1.00	0.00
C	0.00	0.20	-1.00

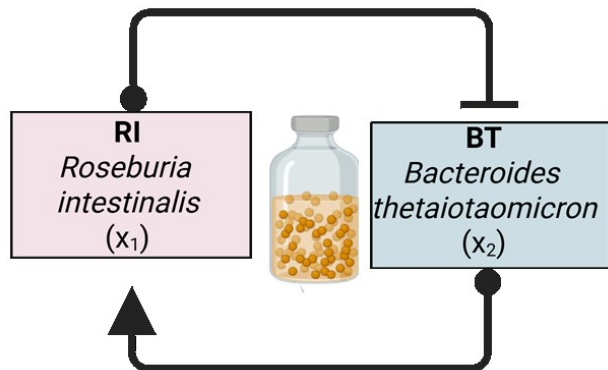
2. By identifying their interactions (and growth rates), we can derive an interactions matrix

3. Allowing us to predict their temporal dynamics

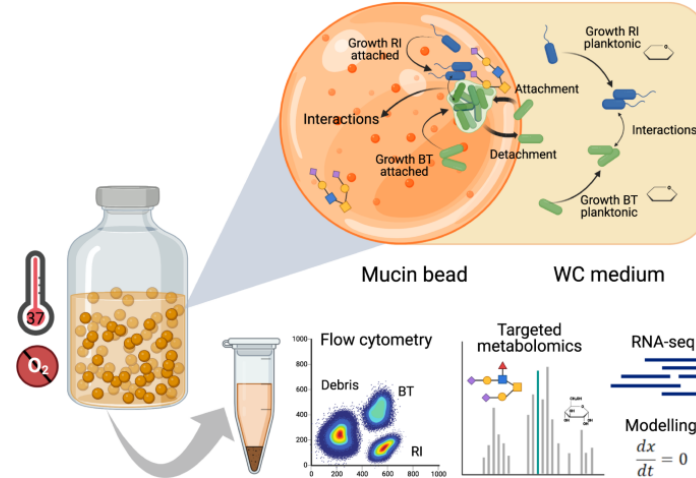
Interactions emerge from local, context-specific dynamics



Initially predicted to compete for pyruvate and glucose



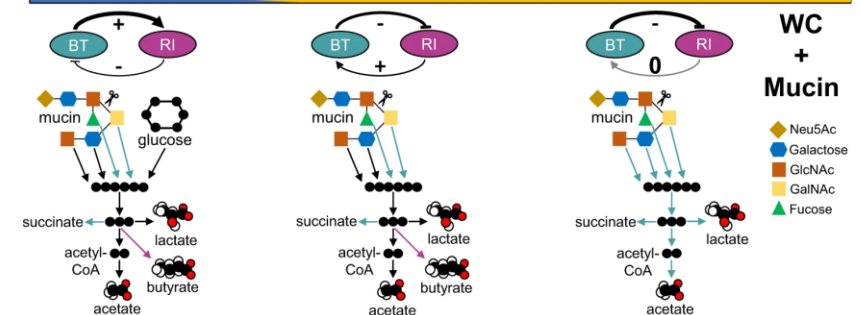
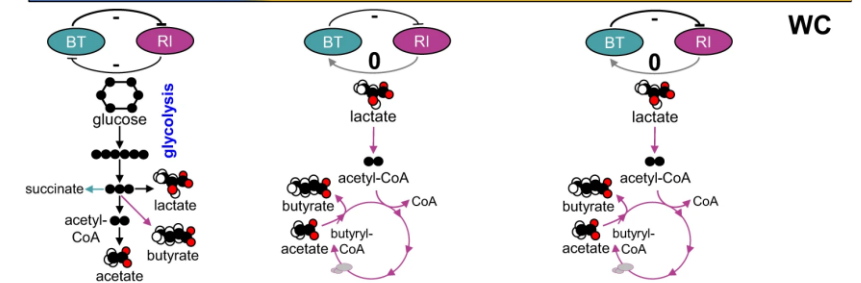
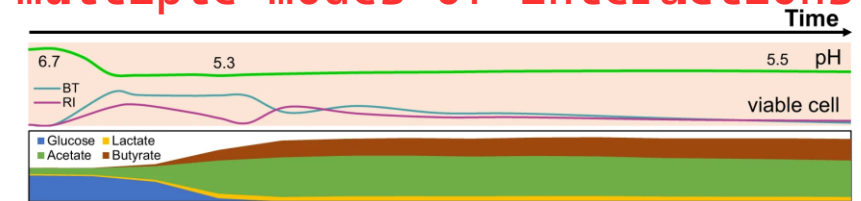
Adding mucin beads is predicted to create positive interactions



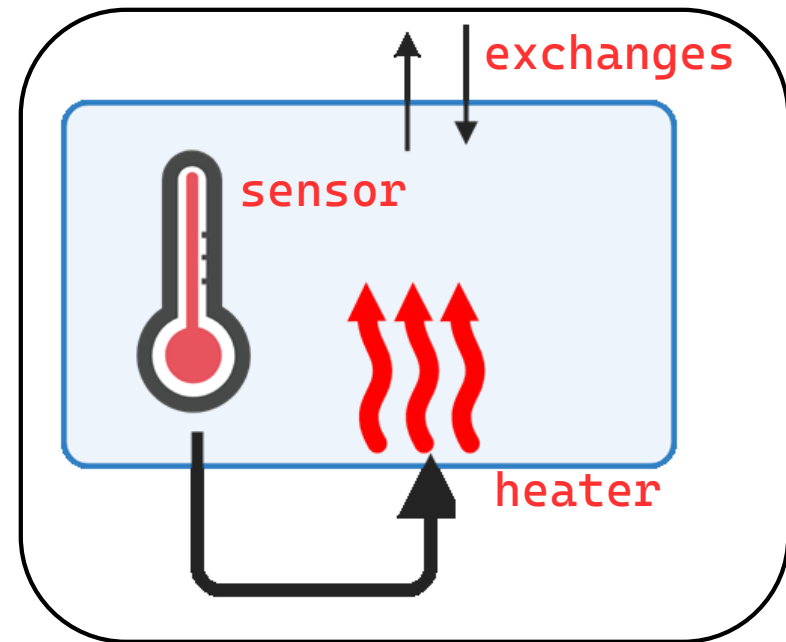
BT Degrades mucin

Both feed on mucin degradation products

Environmental factors lead to multiple modes of interactions



Problem #2: Controlling complex systems requires different control principles



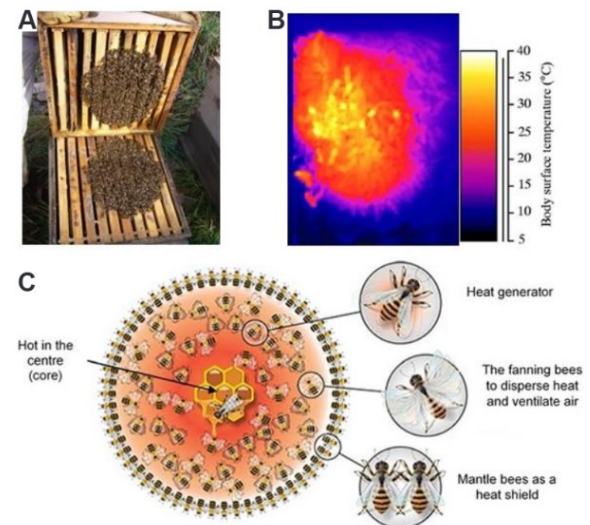
Simple thermostat (outside temperature assumed lower)



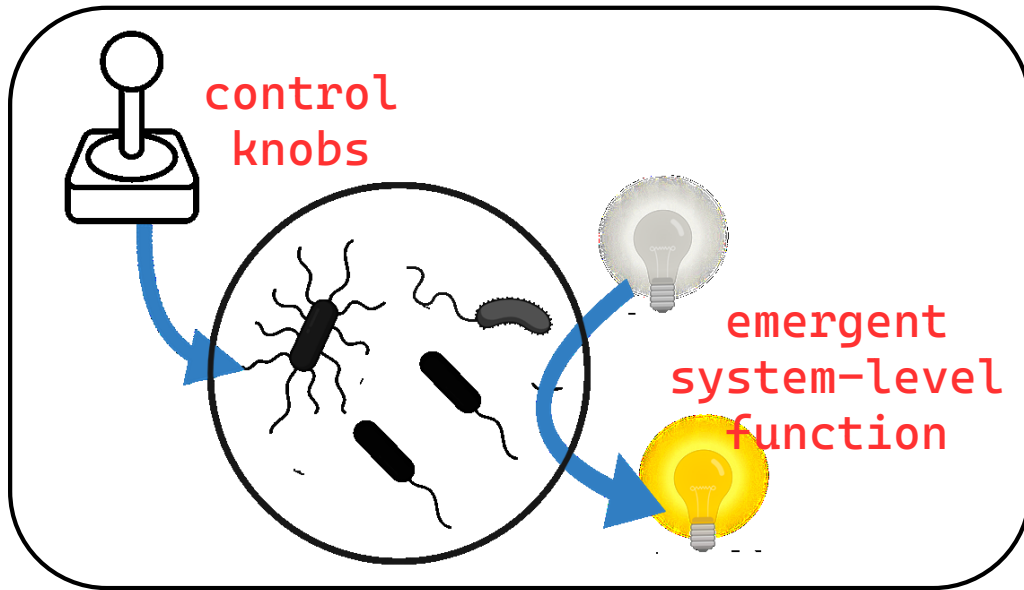
Collective fanning, clustering, and heat sharing maintain brood temperature

Broods need a temperature of 36°C to become healthy adults

Rodriguez-Vasquez et al. 2024



Microbiome engineering is the rational control of microbial ecosystems to make a desired collective function emerge and persist.



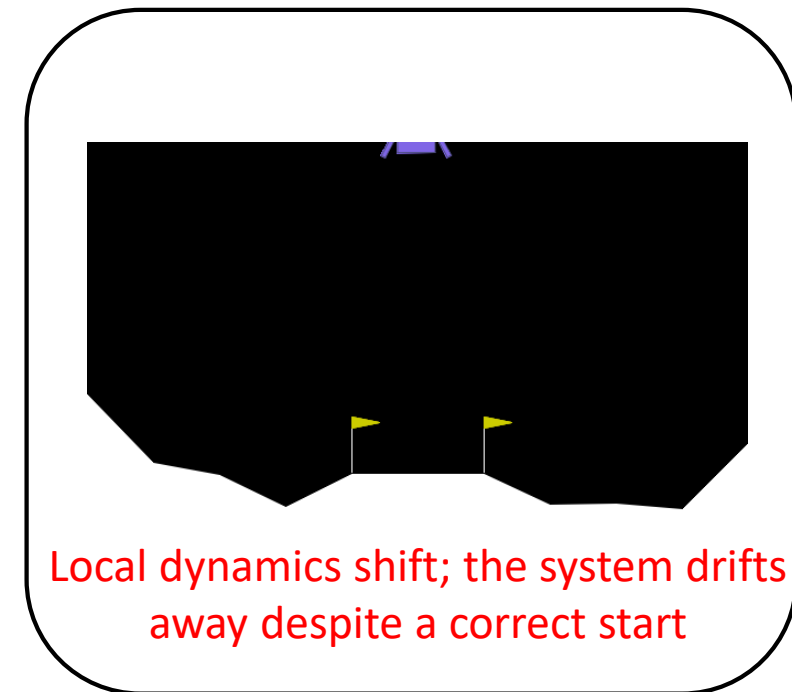
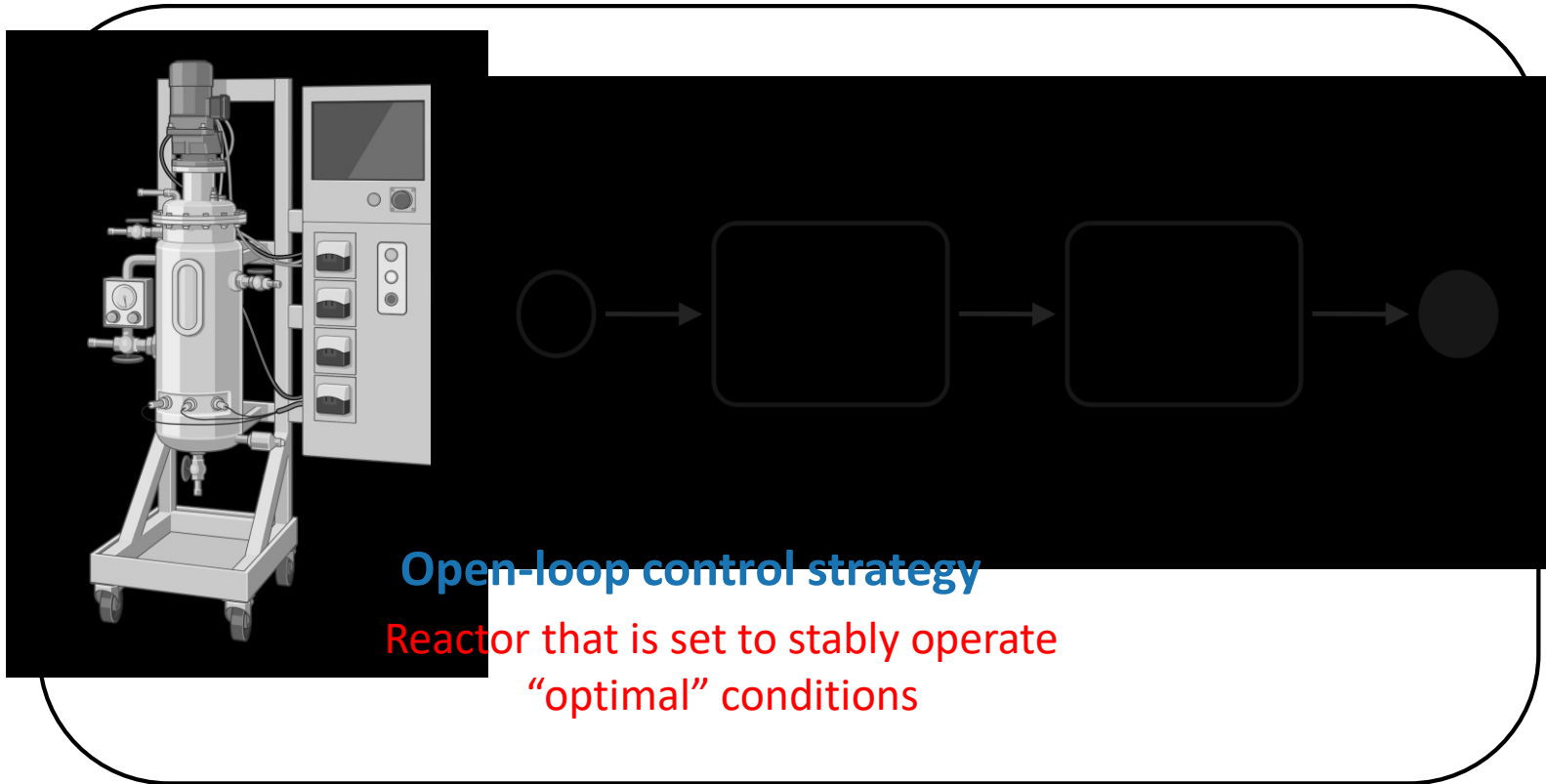
Engineering microbiomes means engineering a complex system.

Complex systems are difficult to predict because their dynamics emerge from local, context-specific interactions.

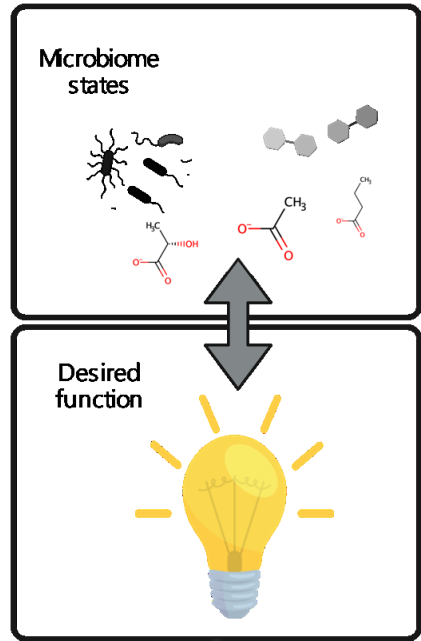
They require fundamentally different control principles.

Most microbiome biotechnologies are insufficient to control complex systems – fundamental ingredients are missing

Missing ingredient #1: feedback control seen from the microbiome's perspective, not just the environment's.

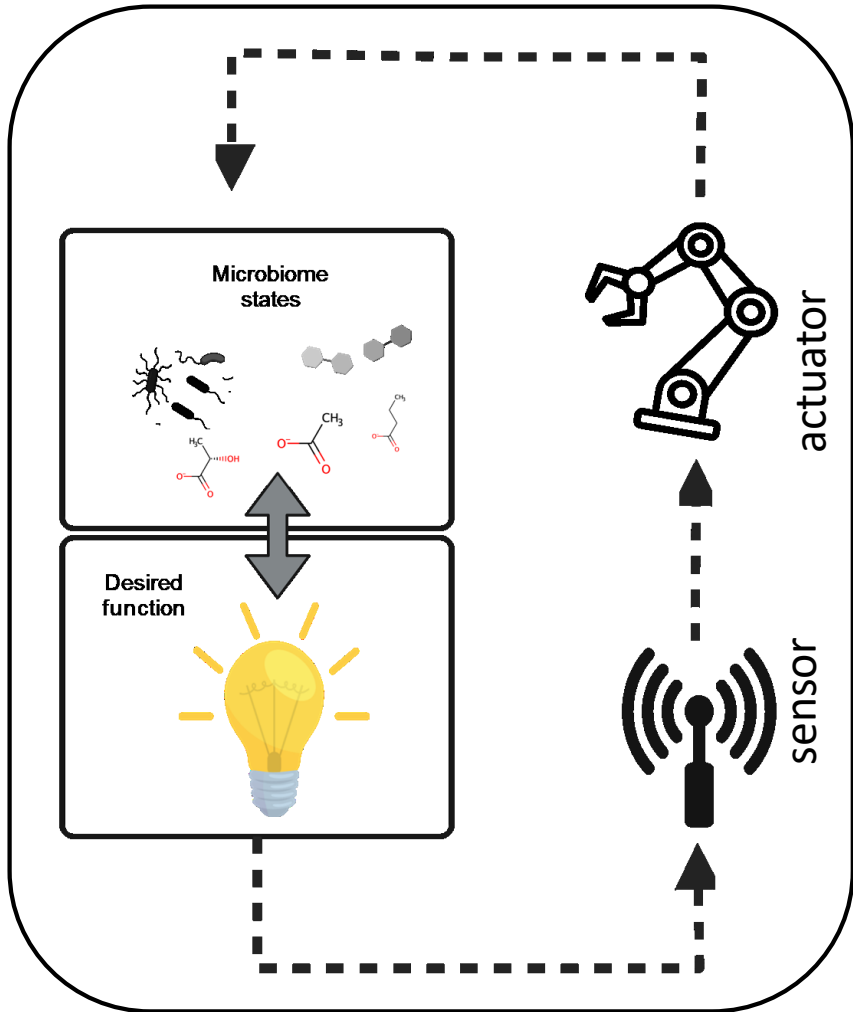


Feedback control means sensing the system and acting to keep it on course



$$X_o \rightarrow (X_o - X_d)$$

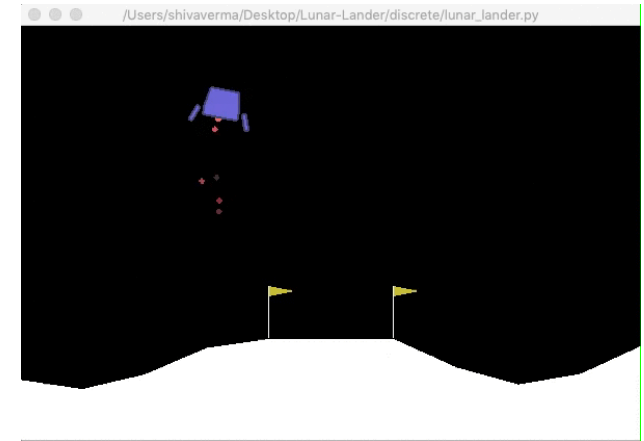
Feedback control means sensing the system and acting to keep it on course



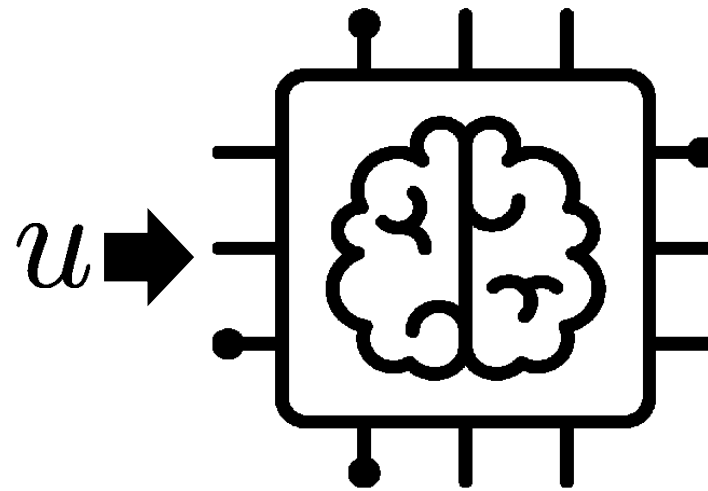
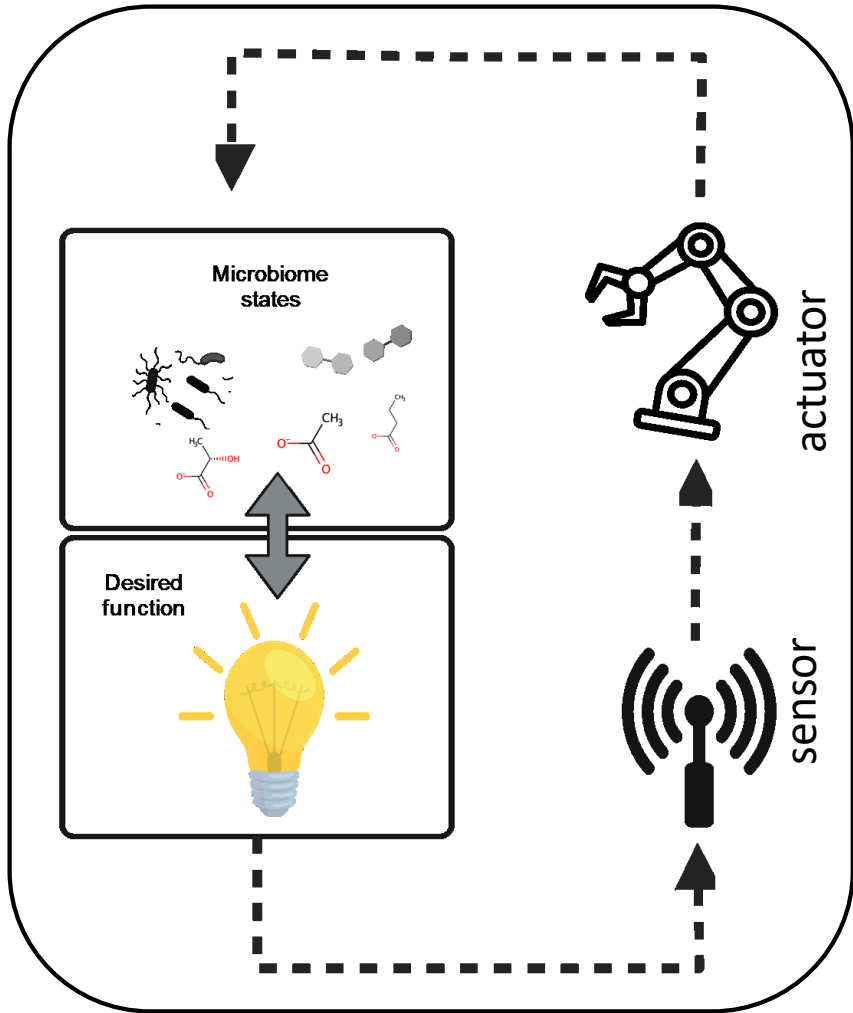
u

$$X_o \Rightarrow (X_o - X_d)$$

Adapts to changes and reaches otherwise unreachable states



Missing ingredient #2: computational intelligence

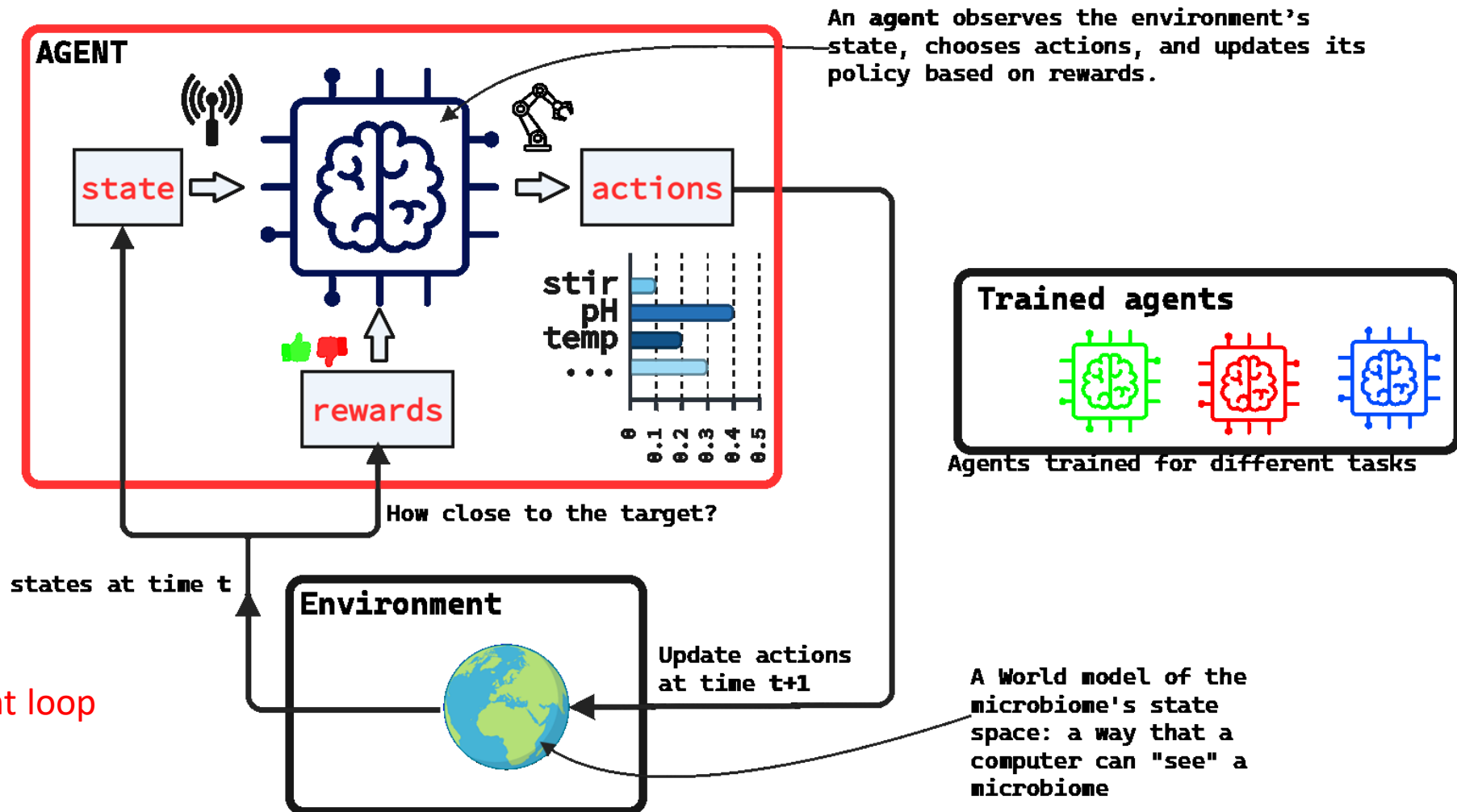


$$X_o \Rightarrow (X_o - X_d)$$

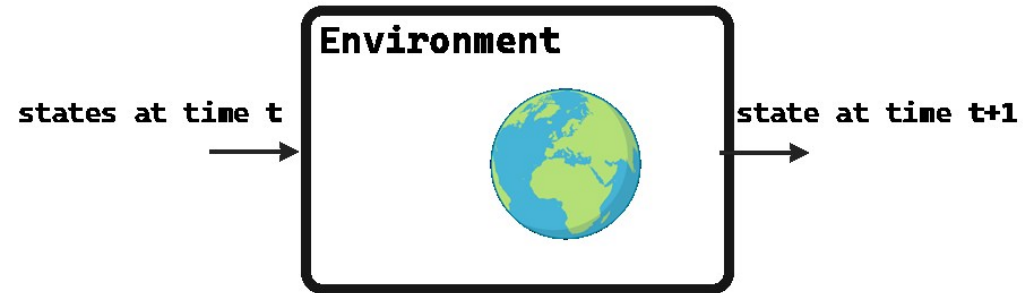
“The global capacity to act purposefully, to think rationally, and to deal effectively with one's environment” – David Weschler

Controlling complex systems by synchronizing local interactions into the correct group-level behavior is a classical problem of intelligence

Computational intelligence is an agent's ability to achieve goals in many environments

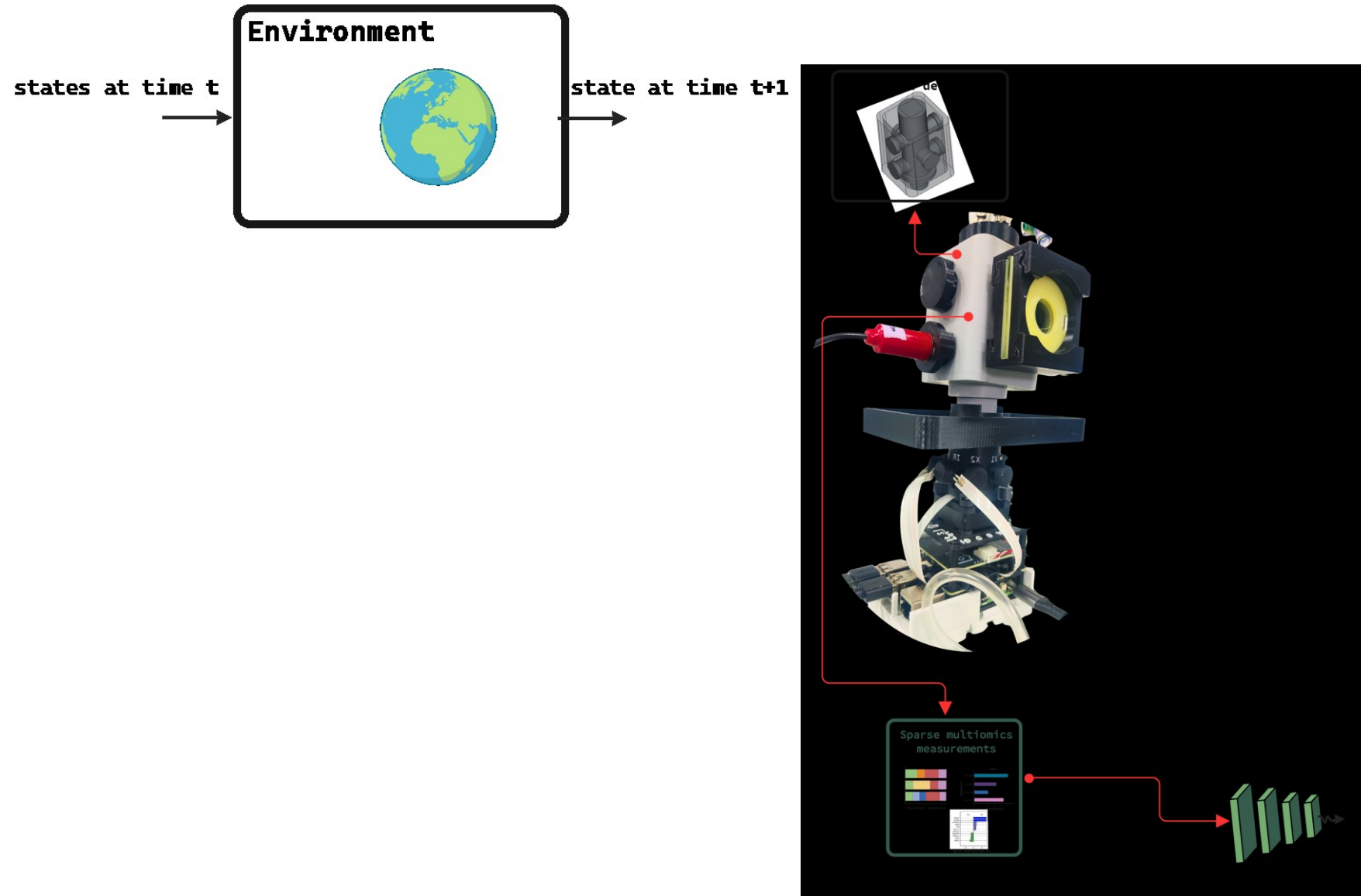


The environment: present the microbiome (physical entity) to the agent (digital entity)

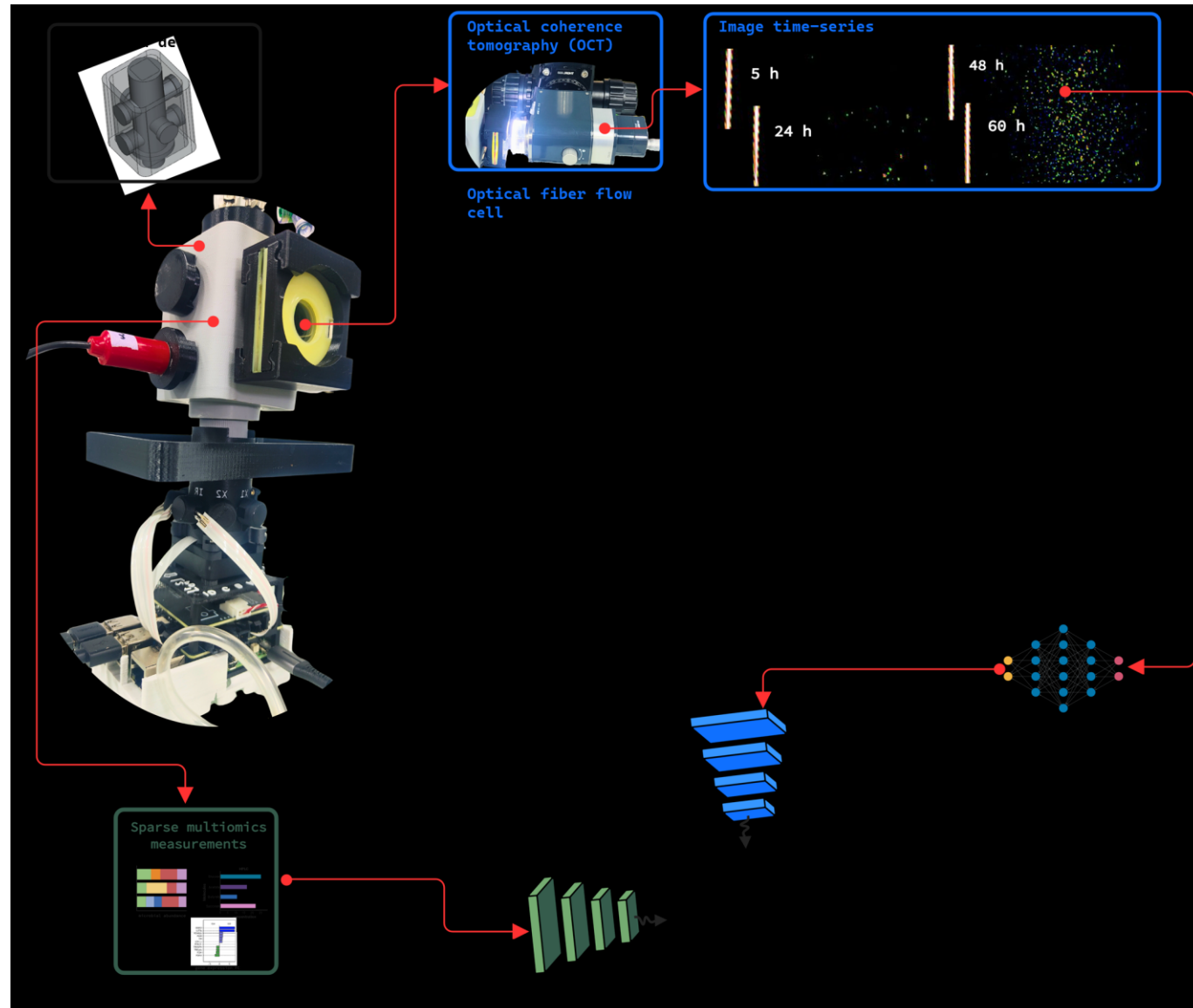


Prototype that directly couples trained agents with living microbiomes to control them in real time.

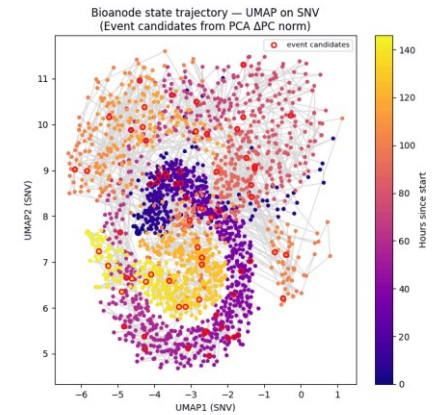
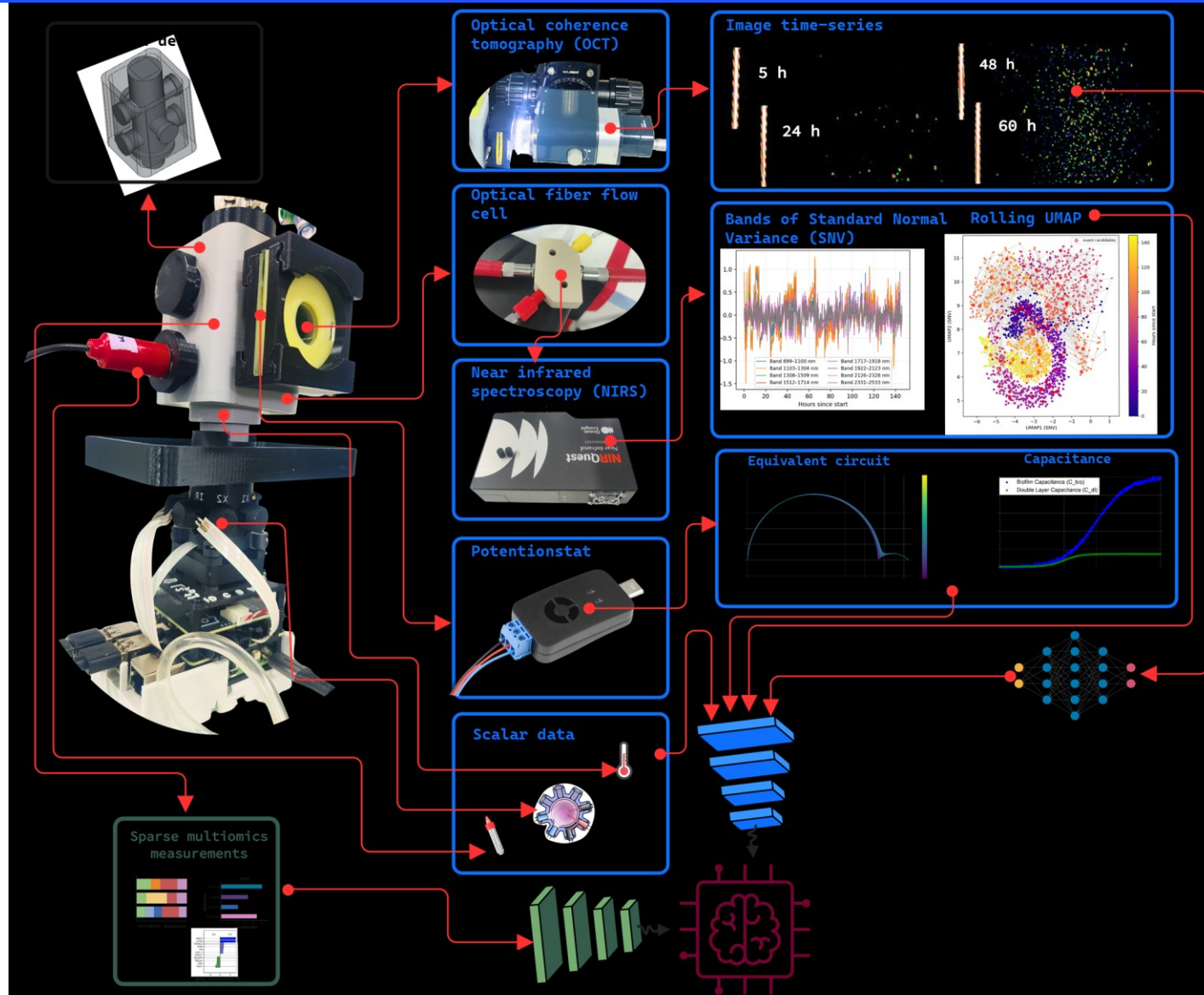
The environment: present the microbiome (physical entity) to the agent (digital entity)



The environment: present the microbiome (physical entity) to the agent (digital entity)

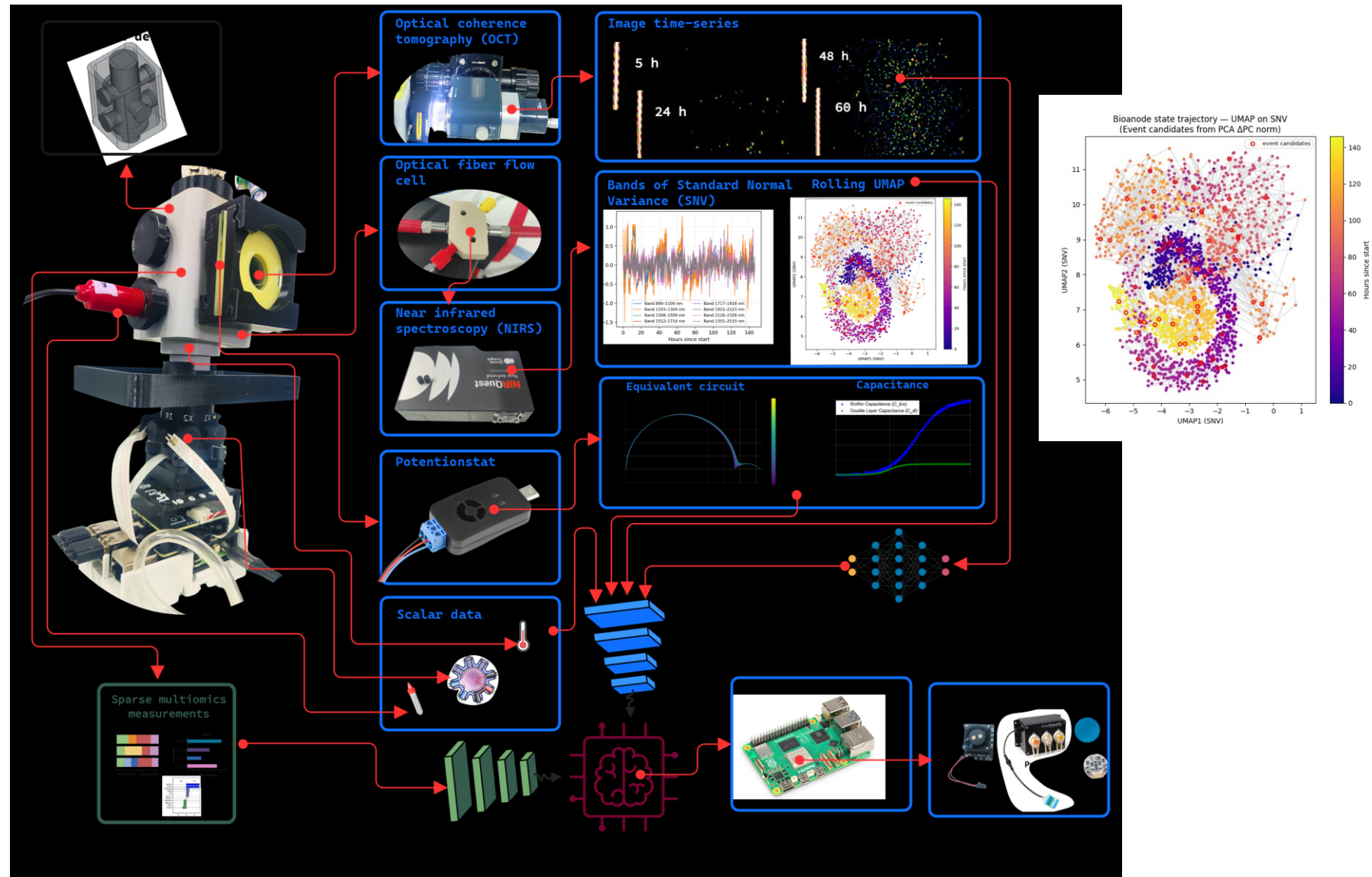


The environment: presentint the microbiome (physical entity) to the agent (digital entity)



The environment: presentint the microbiome (physical entity) to the agent (digital entity)

Prototype that directly couples trained agents with living microbiomes to control them in real time.



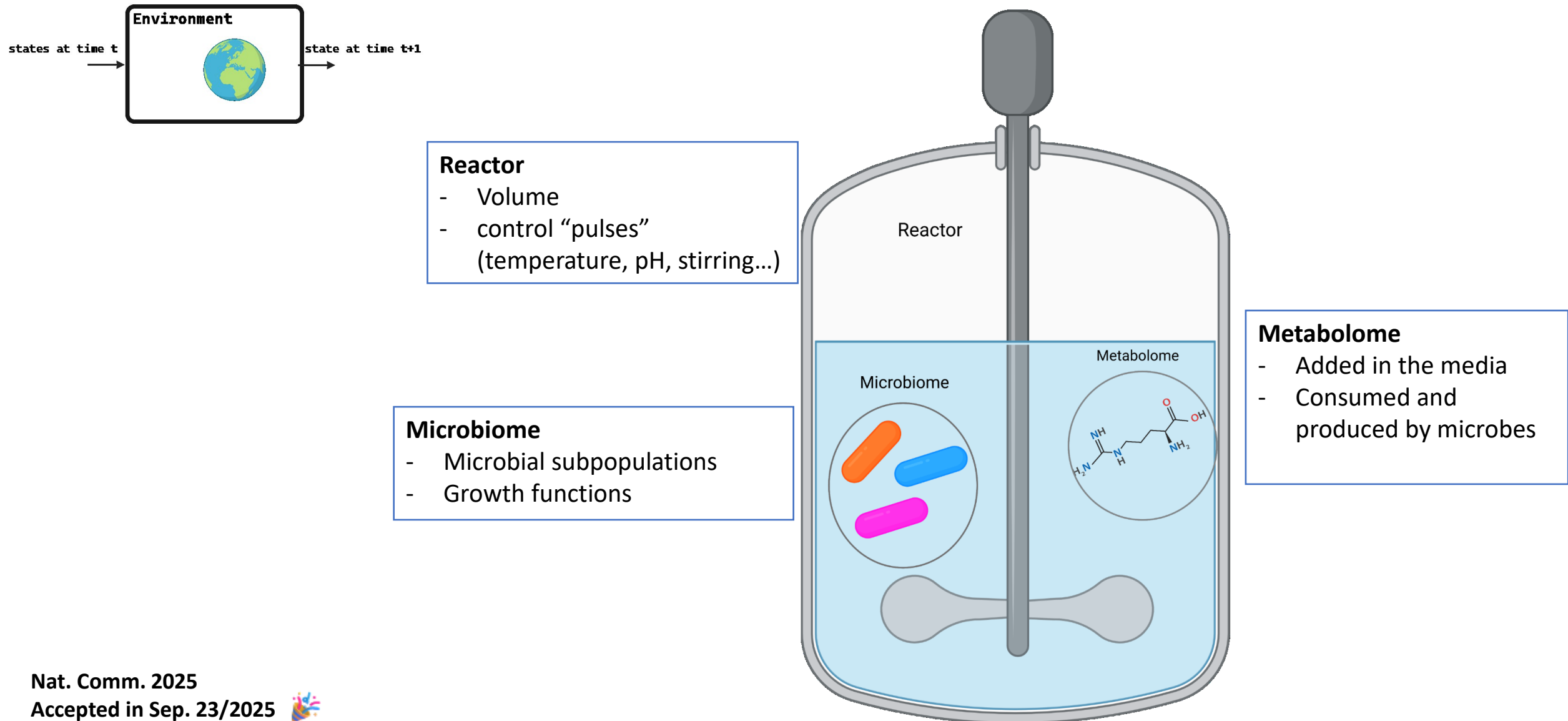
Environment

[illegible]

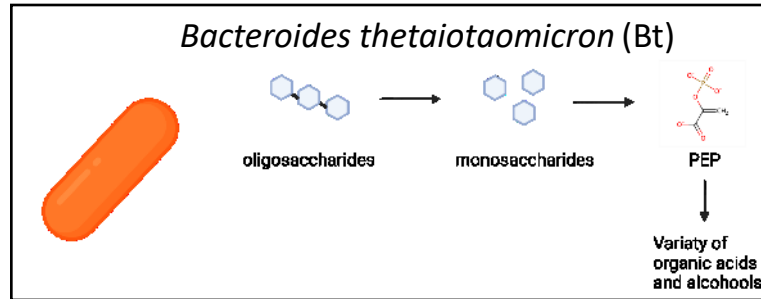
$$\overline{f}(\mathbf{X}, \mathbf{S})$$

2. Formulate mathematical models that enable forward simulation

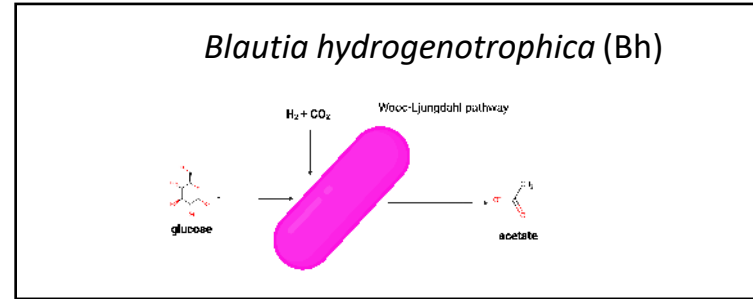
Another way to represent the environment is through a kinetic model



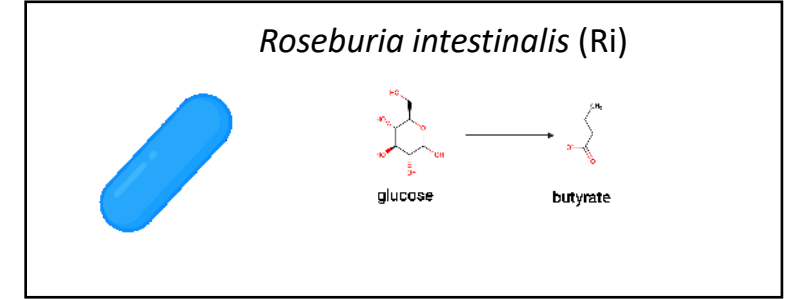
Example of a kinetic model applied to a consortium of three gut bacteria



Primary fermenter



Acetogen



Butyrate producer



live cells

10⁵ cells/uL

Time (h)

Legend:

- bt1 (green circles)
- bt1 simul (black line)
- bt2 (blue circles)
- bt3 (red circles)

Time (h)	bt1 (10 ⁵ cells/uL)	bt1 simul (10 ⁵ cells/uL)	bt2 (10 ⁵ cells/uL)	bt3 (10 ⁵ cells/uL)
0	0.0	0.0	0.0	0.0
10	2.5	2.5	2.5	2.5
20	9.5	9.5	10.5	11.5
30	8.5	8.5	9.5	9.5
40	5.5	5.5	9.5	9.5
50	1.0	4.5	7.5	7.5
60	0.5	2.5	4.5	4.5
70	0.2	1.5	3.5	3.5
80	0.1	0.8	2.5	2.5
90	0.1	0.5	1.5	2.5
100	0.1	0.3	1.0	2.0
110	0.1	0.2	0.5	1.5
120	0.1	0.1	0.2	1.0

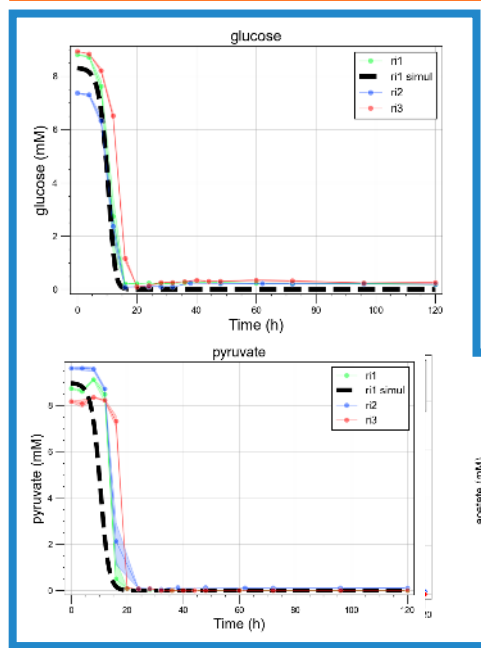
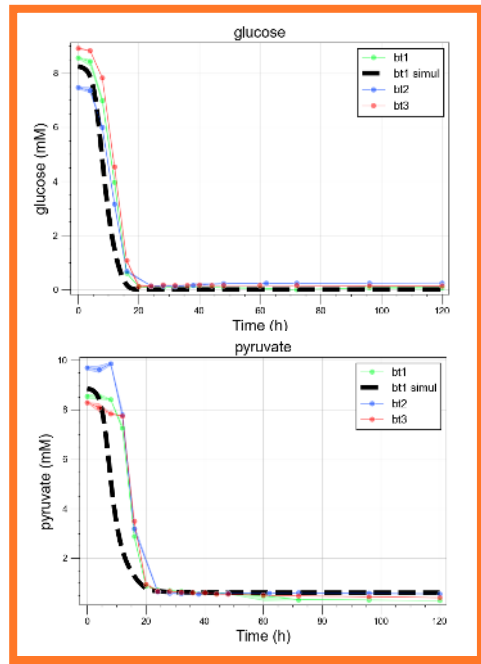
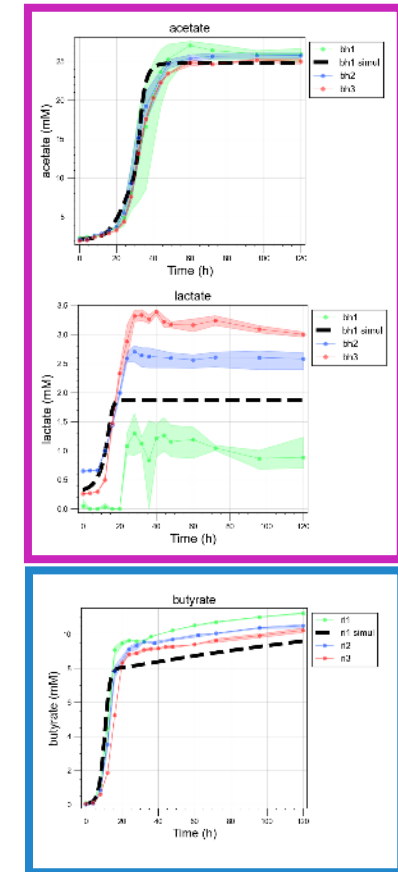


Figure 1 consists of four vertically stacked line graphs showing the time course of metabolite production by *E. coli* strains. The x-axis for all graphs is 'Time (h)' ranging from 0 to 120. The y-axis represents concentration in mM. The legend for all graphs is: t11 (green circles), t11 s/m,1 (black squares), L12 (blue triangles), and t13 (red diamonds). Shaded regions represent standard deviation.

- Formate (mM):** The y-axis ranges from 1.5 to 3.0. All strains show a rapid increase in formate production, reaching a plateau of approximately 2.8 mM by 20-30 hours.
- Lactate (mM):** The y-axis ranges from 1 to 2.5. t11 shows the highest production, reaching a plateau of approximately 2.2 mM. The other strains reach a plateau of approximately 1.8 mM.
- Acetate (mM):** The y-axis ranges from 8 to 14. All strains show a rapid increase in acetate production, reaching a plateau of approximately 12.5 mM by 20-30 hours.
- Succinate (mM):** The y-axis ranges from 0 to 10. All strains show a rapid increase in succinate production, reaching a plateau of approximately 10 mM by 20-30 hours.



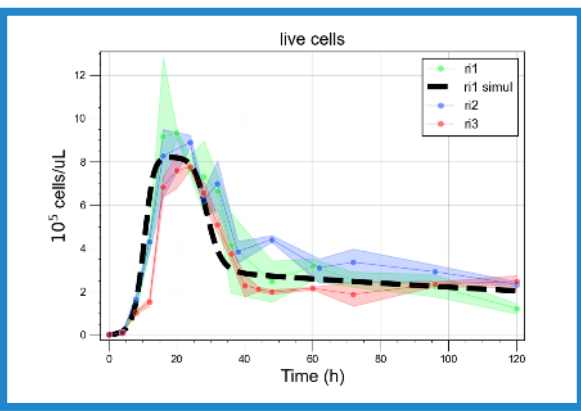
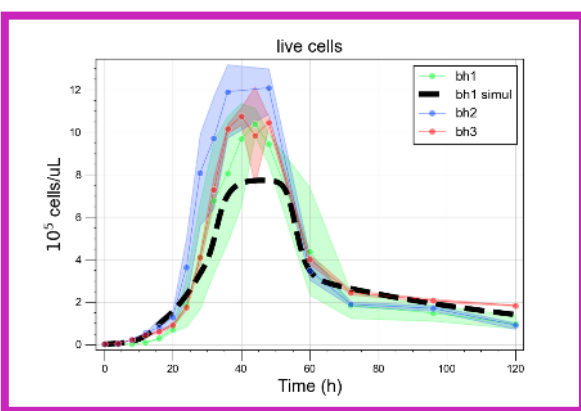
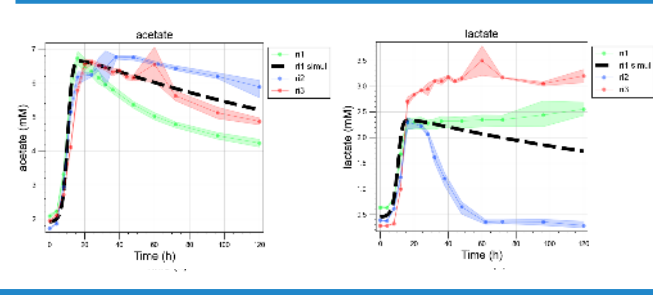
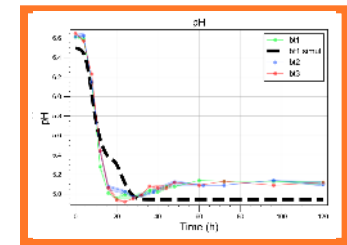
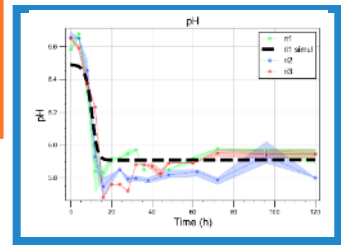
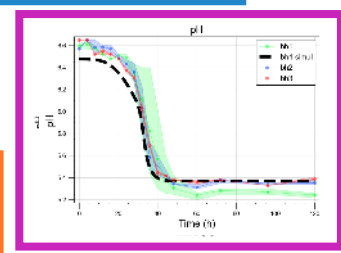
The figure consists of two line graphs showing the pH of a solution over time (0 to 120 hours) during the degradation of an alginate-chitosan hydrogel. The top graph is for pH 1 and the bottom graph is for pH 12. Both graphs show a rapid decrease in pH over time, with experimental data points and a fitted curve.

Top Graph (pH 1):

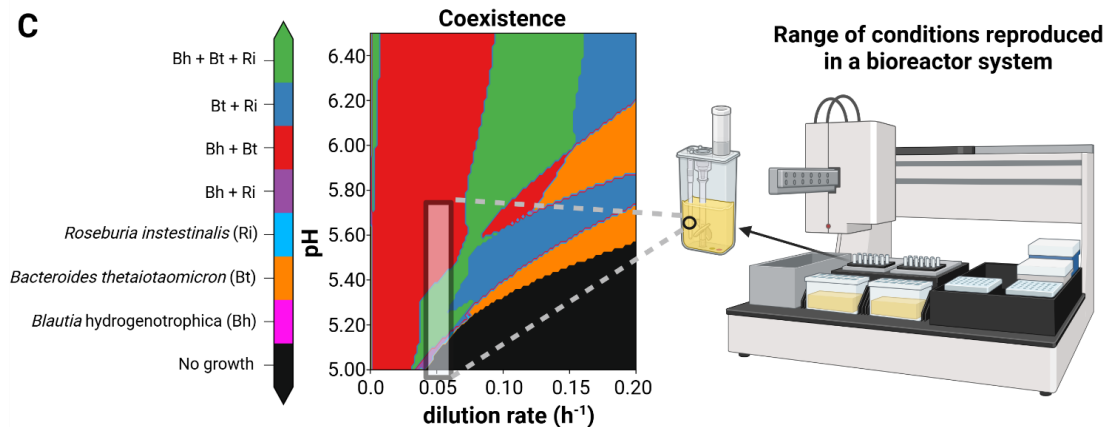
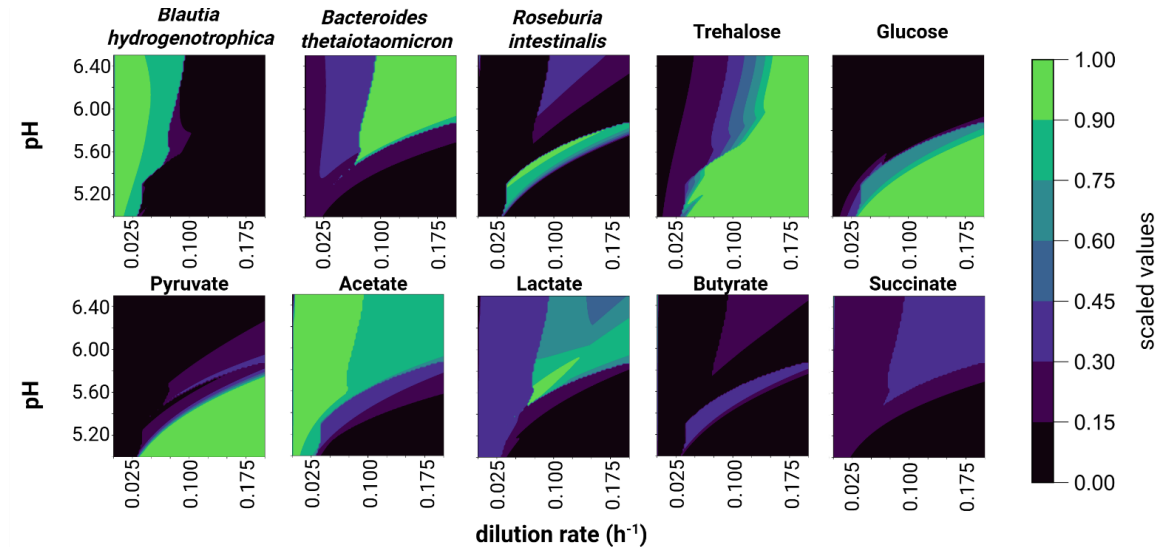
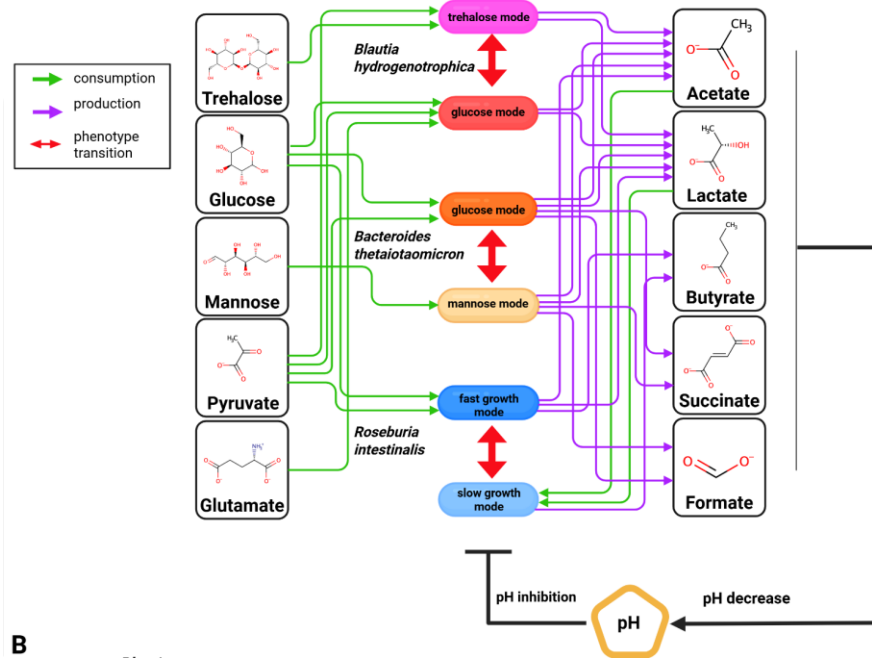
- X-axis:** Time (h) from 0 to 120.
- Y-axis:** pH from 0.7 to 4.0.
- Legend:**
 - bb (black line with dots)
 - bb1 (red line with dots)
 - bb2 (blue line with dots)
 - bb3 (green line with dots)
- Curve:** A dashed black line representing the fitted curve. It starts at pH ~3.8 at 0h, drops sharply to pH ~1.0 by 80h, and then levels off.
- Data Points:** Experimental data points for four different samples (bb, bb1, bb2, bb3) are shown. They all follow a similar trend, dropping from pH ~3.8 to pH ~1.0 by 80h.

Bottom Graph (pH 12):

- X-axis:** Time (h) from 0 to 120.
- Y-axis:** pH from 5.0 to 6.0.
- Legend:**
 - bb (black line with dots)
 - bb1 (red line with dots)
 - bb2 (blue line with dots)
 - bb3 (green line with dots)
- Curve:** A dashed black line representing the fitted curve. It starts at pH ~5.8 at 0h, drops sharply to pH ~5.2 by 20h, and then levels off.
- Data Points:** Experimental data points for four different samples (bb, bb1, bb2, bb3) are shown. They all follow a similar trend, dropping from pH ~5.8 to pH ~5.2 by 20h.

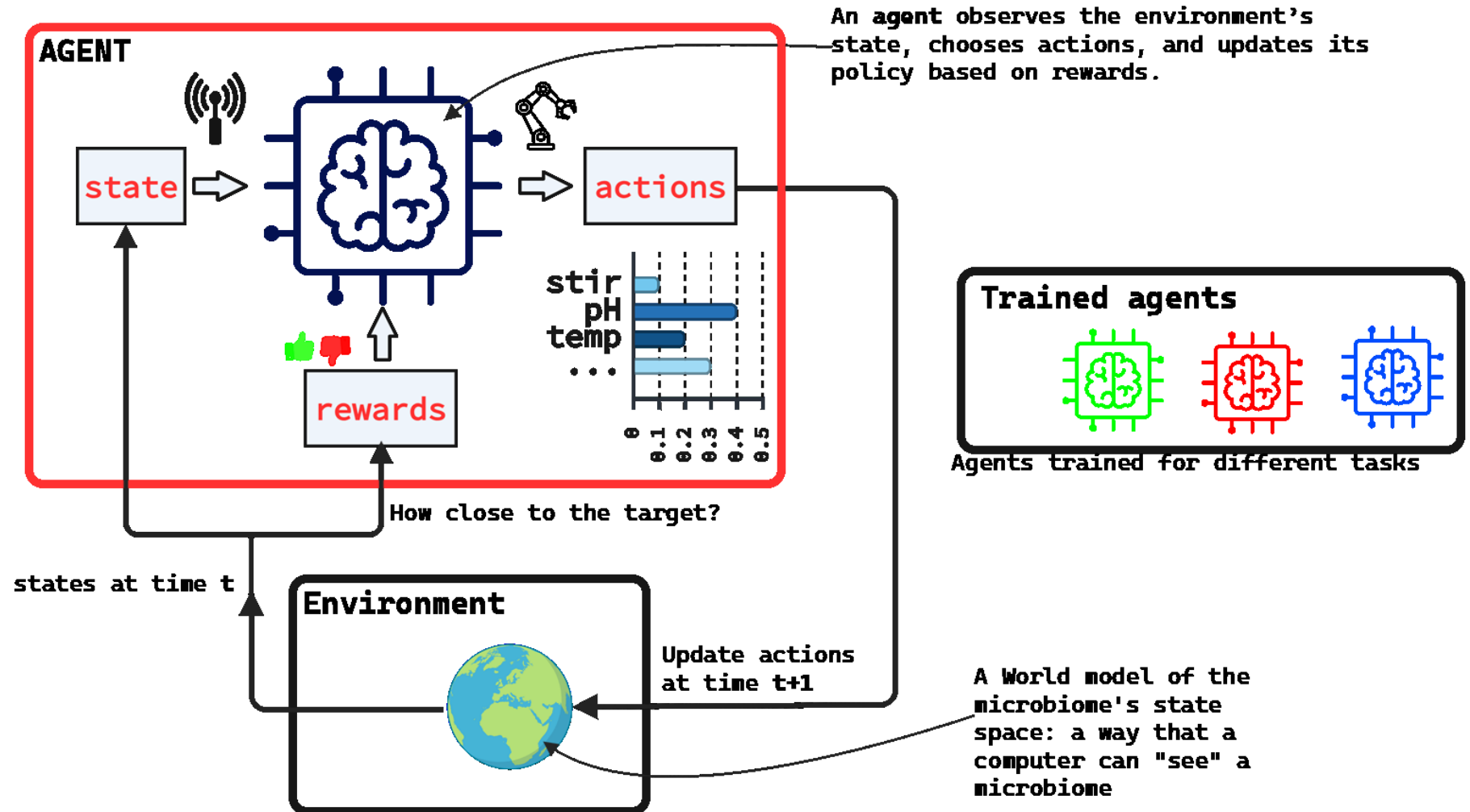


Next we predict the model's phase plane—an open-loop controller

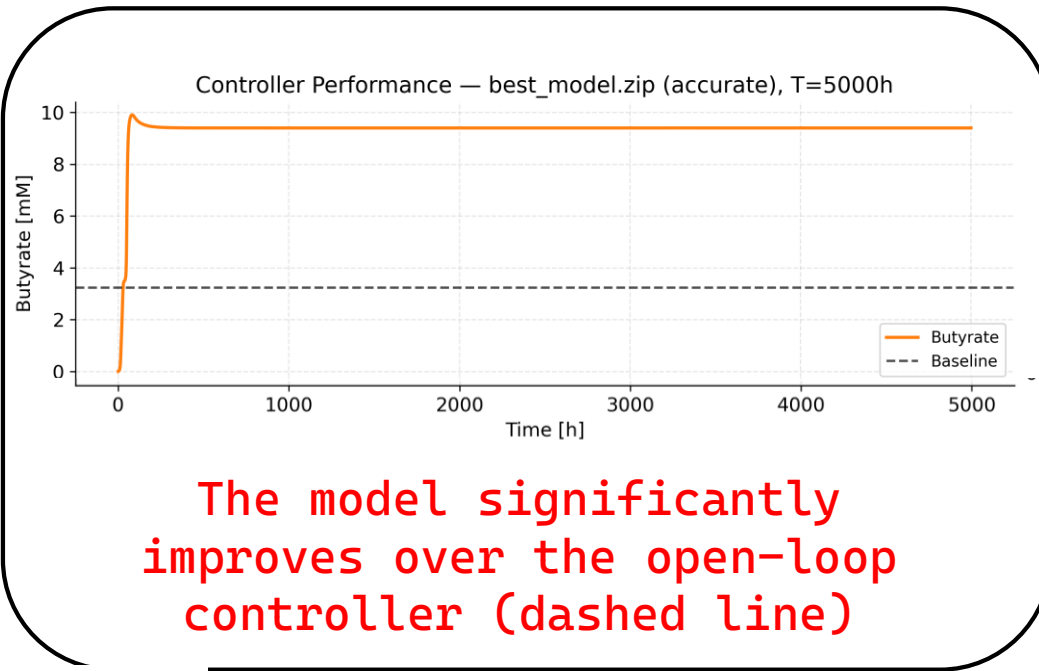


Doing better with an artificially intelligent closed-loop controller?

Agent-Environment loop

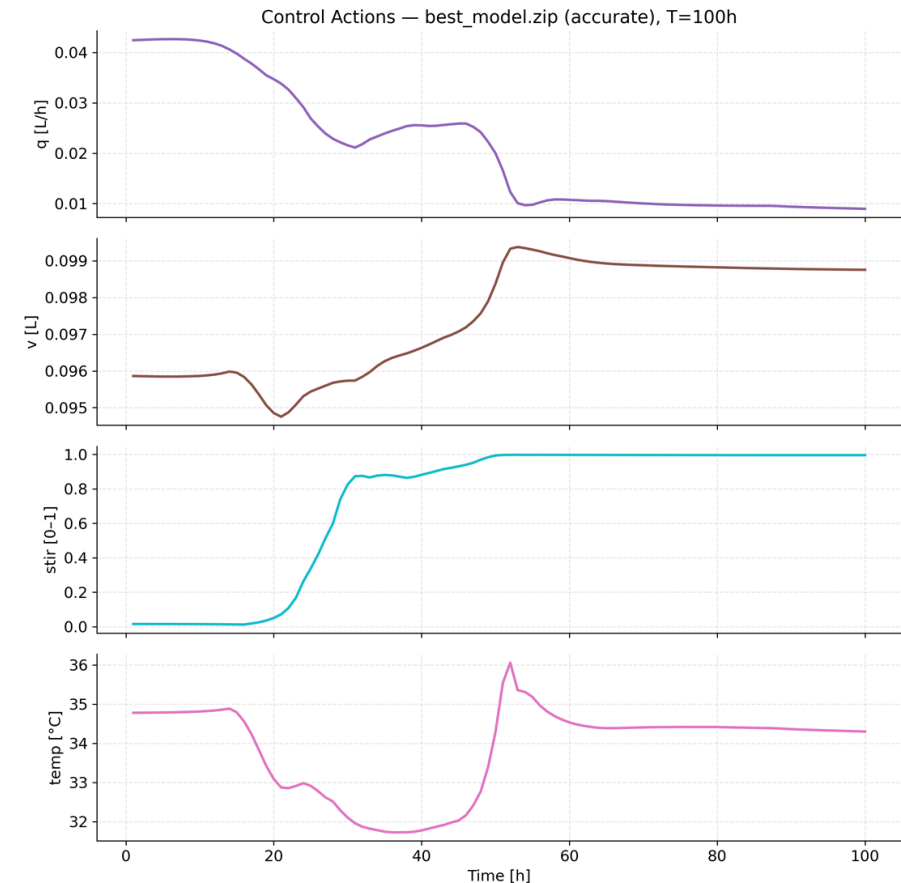


Example: controlling butyrate production



Four actuators:

- 1) Continuous feed.
- 2) Periodic feed (serial dilution).
- 3) Stirring.
- 4) Temperature.

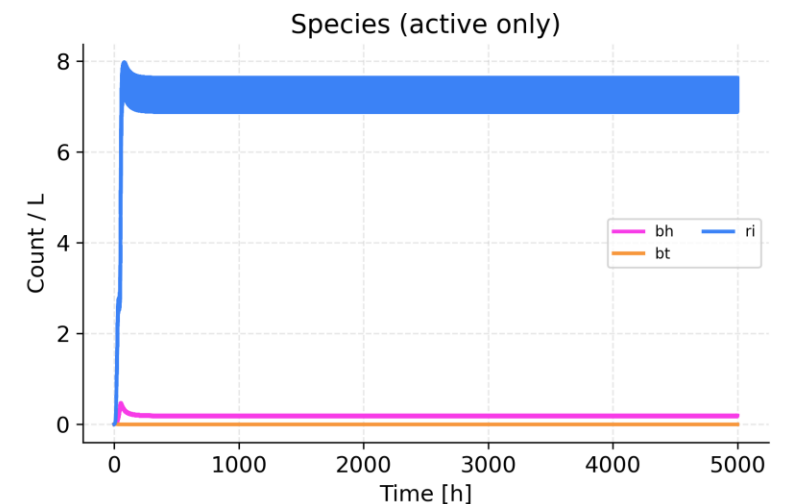
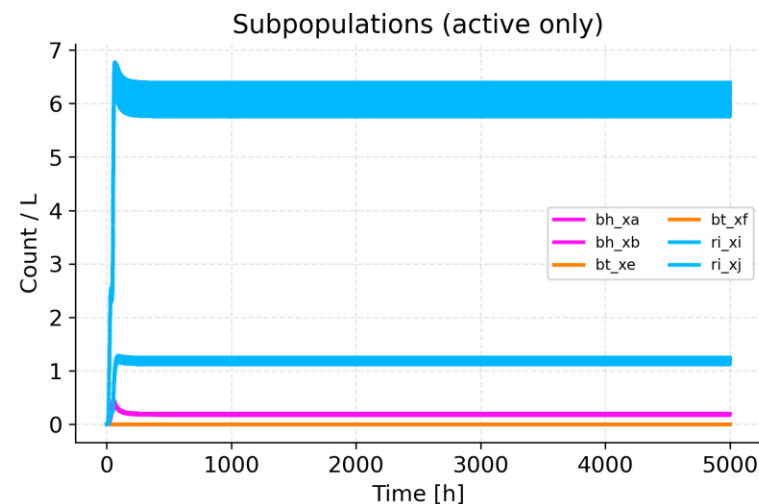
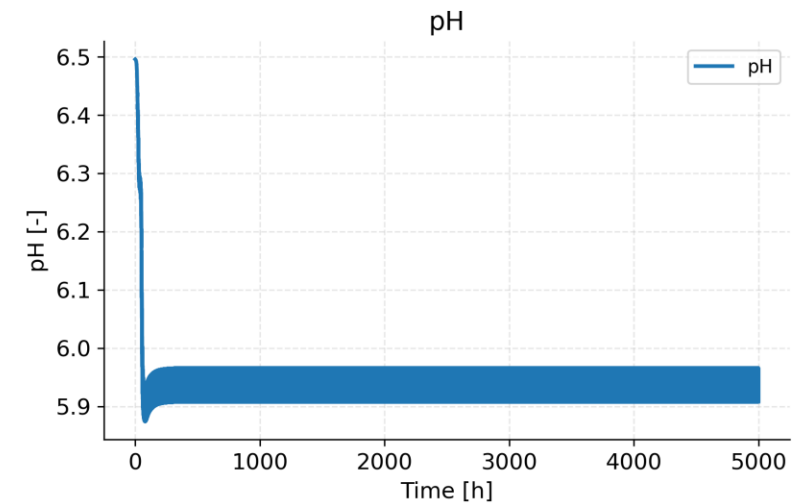
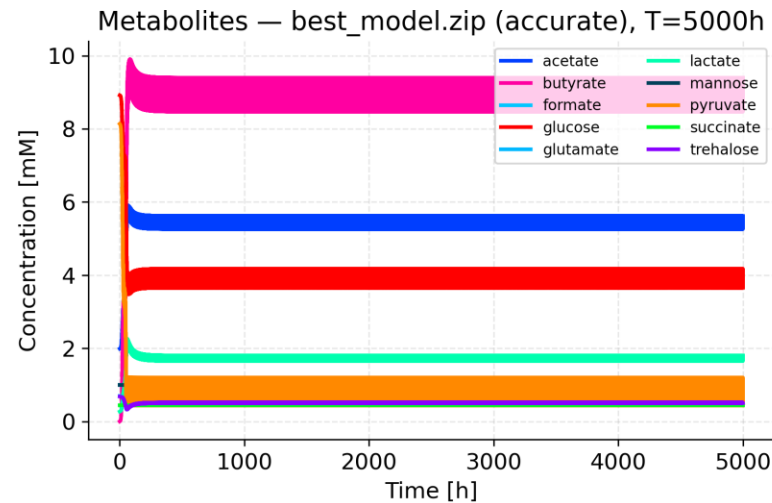


It learns a creative strategy: decreasing continuous feed while increasing periodic feed.

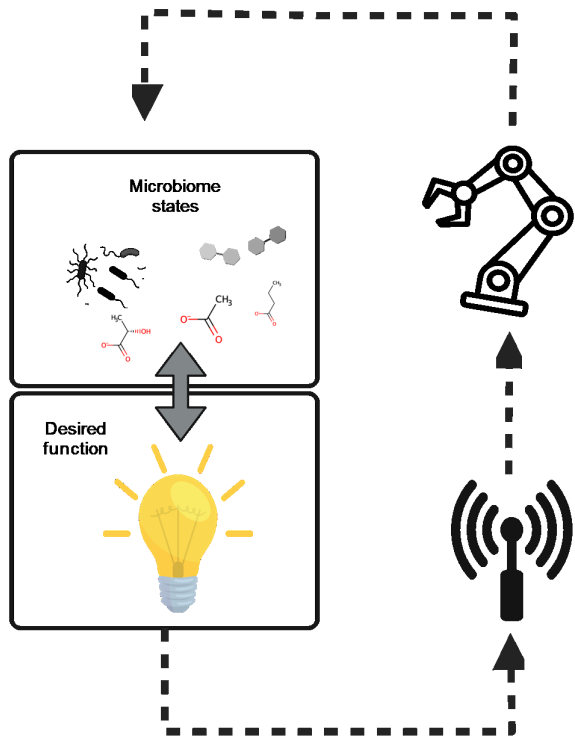
Example: how butyrate is maximized

The solution keeps a small population of the acetogen (*Blautia hydrogenotrophica*) feeding on trehalose, while producing acetate that the butyrate producer (*Roseburia intestinalis*) can use to make more butyrate.

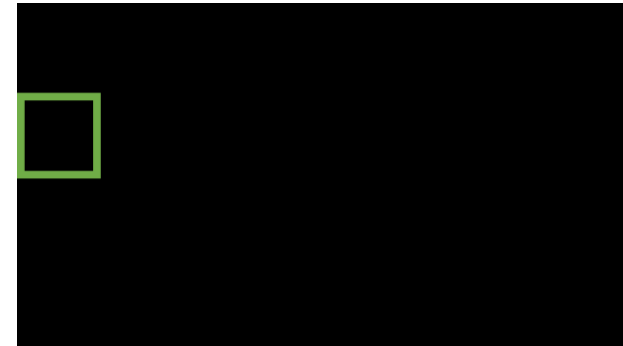
When feeding on trehalose, the acetogen does not compete for the other carbon sources (glucose and pyruvate)



Summary



- 1) Microbiomes are complex system that are difficult to predict and control
- 2) We need intelligent feedback controllers to move them towards desired states, which are often far from their natural equilibrium.
- 3) Essentially, the challenge of closed-loop microbiome control is the challenge of developing intelligent agents capable of interpreting the current state and finding actions to move the system towards desired states.



Practical task: Use the kinetic model to compare a random controller with a trained controller